

Leica CM1860/ Leica CM1860 UV

Cryostat

((

Instructions for Use

Leica CM1860/CM1860 UV, V1.4 – 10/2012, English Order No.: 14 0491 80101 RevE

Always keep this manual with the instrument. Read carefully before working with the instrument.



The information, numerical data, notes and value judgments contained in this manual represent the current state of scientific knowledge and state-of-the-art technology as we understand it following thorough investigation in this field. We are under no obligation to update the present manual periodically and on an ongoing basis according to the latest technical developments, nor to provide our customers with additional copies, updates etc. of this manual.

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For the instrument serial number and year of manufacture, please refer to the nameplate on the back of the instrument.

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Important Information

1.1 Symbols in the text and their meanings



Warnings

appear in a gray box and are marked by a warning triangle



Useful tips.

i.e. important information for the user. appear in gray boxes and are marked by an 🚦 .



Caution – UVC radiation!

(5) (Fig.5)

Numbers in parentheses refer to item numbers in illustrations or to the illustrations themselves.



Manufacturer



This product fulfills the requirements of the Council's Directive 98/79/EC concerning in vitro diagnostics (IVD) medical devices.



In vitro diagnostics (IVD) medical device



Observe the Instructions for Use

REF Order No.

SN Serial number



Symbol for labeling electrical and electronic equipment in accordance with Section 7 of the German Electrical and Electronic Equipment Act (ElektroG). ElektroG is the law governing how electrical and electronic equipment is brought into circulation, returned and disposed of in an environmentally friendly manner.



The CSA test mark means that a product has been tested and fulfills the applicable safety and/or performance standards, including the relevant standards defined or administered by the American National Standards Institute (ANSI), Underwriters Laboratories (UL), the Canadian Standards Association (CSA), the National Sanitation Foundation International (NSF) and others.

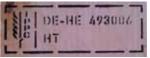


Environmental protection symbol of the China RoHS directive. The number in the symbol indicates the "Environmentfriendly Use Period" of the product. The symbol is used if a substance restricted in China is used in excess of the maximum permitted limit.

Important Information



The package contents are fragile and must be handled with care.



Example of labeling in accordance with **IPPC**



Indicates the correct upright position of the package.



The package must be kept in a dry environment.



It is not permitted to stack packages, and no loads may be placed on top of the package.

Storage temperature range:



Indicates the temperature range permitted for storing and handling the package.

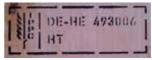
Minimum +5°C Maximum +50°C

Transport temperature range



Indicates the temperature range permitted for transporting the package.

Minimum -40°C Maximum +55°C



- IPPC symbol
- Country code in accordance with ISO 3166, e.g. DE for Germany
- Regional identifier, e.g. HE for Hessen
- Producer/treatment provider code, unique assigned number starting with 49
- Treatment code, e.g. HT (heat treatment), MB (methyl bromide), and possibly DB (debarked)



Package labeling in accordance with German Hazardous Freight Ordinance Road and Rail (GGVSE)/European Agreement concerning the International Carriage of Dangerous Goods by Road (ADR) for transporting hazardous goods. Class 3: "FLAMMABLE LIQ-UID"



Tip-n-Tell indicator to monitor whether the shipment has been transported and stored in upright position according to your requirements. With a pitch of 60° or more, the blue quartz sand flows into the arrow-shaped indicator window and sticks there permanently. Improper handling of the shipment is immediately detectable and can be proven definitively.

1. Important Information

1.2 Qualification of personnel

Only trained laboratory personnel may operate the Leica CM1860/CM1860 UV. Prior to operating the instrument, the operator must thoroughly read and understand these Instructions for Use and must familiarize him/herself with all technical details of the instrument.



Despite chemical and/or UV-light disinfection, adequate safety measures as per the applicable laboratory regulations must still be taken (i.e. safety goggles, gloves, laboratory coat and mask must be worn). These disinfections mean a reduction in the number of germs of at least 99.99% (see certificate on the Leica website).

1.3 Intended use of instrument

The Leica CM1860/CM1860UV is a high-performance cryostat with disinfection system for routine and research applications in biology, medicine and industry. The instrument is used to freeze and section specimen material quickly. The instrument is not designed for unsupervised storage of specimen material. The instrument may only be operated within the scope of its designated use as described above and as per the instructions given in these Instructions for Use. The Leica CM1860/CM1860UV is also suitable for IVD (in vitro diagnostics).

Any other use of the instrument is considered improper.

1.4 Instrument type

All information contained in these Instructions for Use applies exclusively to the instrument type indicated on the title page. A nameplate bearing the instrument serial number (SN) is attached to the back of the instrument.

These Instructions for Use include important instructions and information related to the operating safety and maintenance of the instrument. The Instructions for Use are an important part of the product, and must be read carefully prior to startup and use and must always be kept near the instrument.

If additional requirements on accident prevention and environmental protection apply in the country of operation, these Instructions for Use must be supplemented by appropriate instructions to ensure compliance with such requirements.

Proper and intended use includes the observance of all instructions in the Instructions for Use and compliance with all inspection and maintenance instructions.

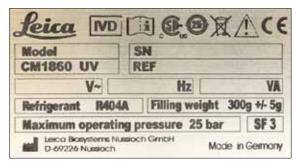


Fig. 1

The nameplate shown here is intended as an example only!



The safety and caution notes in this chapter must be observed at all times. Be sure to read these notes even if you are already familiar with the operation and use of other Leica products.

2.1 Safety notes

These Instructions for Use include important instructions and information related to the operating safety and maintenance of the instrument.

The Instructions for Use are an important part of the product, and must be read carefully prior to startup and use and must always be kept near the instrument.

This instrument has been built and tested in accordance with the safety requirements for electrical equipment for measurement, control, and laboratory use.

Residual risks

The instrument has been designed and constructed with the latest state-of-the-art technology and

according to recognized standards and regulations with regard to safety technology. Operating or handling the instrument incorrectly can place the user or other personnel at risk of injury or can cause damage to the instrument or other property. The machine may be used only as intended and only if all of its safety features are in proper working condition. Malfunctions that impede safety must be remedied immediately.

To maintain this condition and ensure safe operation, the user must observe all notes and warnings contained in these Instructions for Use.

Only original spare parts and permitted original accessories may be used.



For the CE certificate and up-todate certificates pertaining to UV disinfection, visit us online at: www.LeicaBiosystems.com



These Instructions for Use must be appropriately supplemented as required by the existing regulations on accident prevention and environmental safety in the operator's country.



The protective devices on both instrument and components must neither be removed nor modified. Only service personnel qualified by Leica may repair the instrument and access the instrument's internal components.

2.2 Warnings

The safety devices installed in this instrument by the manufacturer only constitute the basis for accident prevention. Operating the instrument safely is, above all, the responsibility of the owner, as well as the designated personnel who operate, service or repair the instrument.

To ensure trouble-free operation of the instrument, make sure to comply with the following instructions and warnings.

2.3 Safety devices

The instrument is equipped with the following safety devices: Handwheel lock and knife guard on the knife holder.



To prevent adverse health effects from UV radiation, the UV disinfection cycle can be started only after the sliding window has been properly closed. Closing the window activates the corresponding safety features.

The consistent use of these safety features and strict observation of the warnings and cautions in these Instructions for Use will safeguard the operator from accidents and/or personal injury to a great extent.

Personal safety precautions



When working with cryostats, personal safety precautions must always be taken. It is mandatory to wear work safety shoes, safety gloves, a mask and safety goggles.



UVC disinfection is effective when disinfecting surfaces and air within the irradiated working area of the Leica CM1850UV, CM1860UV and CM1950 cryostats at -20°C (Table 1, see I. Maier certificate). For powerful disinfection, we recommend irradiation for three hours (CM1850UV/CM1860UV/CM1950). Vegetative bacteria including Mycobacterium tuberculosis, bacterial endospores (Bacillus sp.) and fungi are killed during this time. Even viruses including such resistant species as hepatitis are inactivated up to at least 4 log10 units (99.99%).

Personal safety precautions (continued)



Medium disinfection can be attained through short irradiation for 30 minutes (CM1850UV/CM1860UV/CM1950). This reduces vegetative bacteria including Mycobacterium tuberculosis and sensitive viruses such as the influenza A virus (also including the highly pathogenic avian influenza virus H5N1) and the poliovirus by at least 5 Log 10 units (99.999%). UVC irradiation within the working area of the cryostats can provide reliable and efficient disinfection of surfaces and the air and significantly reduces the risk of infection.

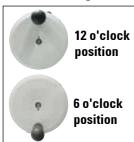
We recommend removing tools and debris and then wiping off visible contamination in the cryostat with an alcohol-based disinfectant prior to using the UV lamps. The germicidal effect of the irradiation is restricted to the directly irradiated areas, which is why UVC irradiation cannot be a replacement for regular chemical disinfection of the cryostat chamber.



For current information about certificates and recommendations, visit:

http://www.LeicaBiosystems.com/products/total-histology/cryo-sectioning/details/product/leica-CM1860-uv/downloads/

2.3.1 Locking/unlocking the handwheel





Rotate the handwheel only if the refrigeration system is on and the cryochamber is cold.



Always lock the handwheel prior to making modifications to the knife or specimen, changing the specimen, or taking a break.

To lock the handwheel, move its handle to the 12 or 6 o'clock position. Press the lever (1) fully outward; gently rock the handwheel back and forth until the locking mechanism clicks into place noticeably.

To release the handwheel, press the lever (2) on the handwheel toward the cryostat housing.





Fig. 3

2.3.2 Knife guard



Prior to manipulating the knife and specimen, changing the specimen or knife, or taking a break, always lock the handwheel and cover the cutting edge with the knife guard.

The CN and CN-Z knife holders feature knife guards; the glass anti-roll plate of the CE and CE-TC blade holders also serves as a knife guard.

2.4 Operating conditions

Transport and installation

- After transporting, do not turn the instrument on for a minimum of 4 hours.
- Do not operate in rooms with explosion hazard!
- To ensure proper function, the instrument must be set up with the following minimum clearance to walls and fixtures on all sides:

- rear: 15 cm - 5.9 inch - right: 30 cm - 11.8 inch - left: 15 cm - 5.9 inch

Electrical connection

- Before connecting the instrument to an AC power outlet, ensure that the AC power of your laboratory matches the values on the nameplate of the instrument.
- During the start-up of the refrigeration system, the minimum nominal voltage must not drop below the values specified in Chapter 3, "Technical Data".

The compressor draws a starting current between 45 and 50 A.

Therefore, the electric circuit at the place of installation must be inspected by an electrical engineer to ensure that it meets the requirements for a smooth operation of the instrument.

A stable power supply in accordance with the instrument's specifications is essential to its proper functioning.

Failure to comply with the above can damage the instrument.

- This waiting period is necessary to allow the compressor oil, which may have been displaced during transport, to return to its original position.
 Failure to comply with this may cause damage to the instrument.
- No additional extension cable may be used.

Defrosting

The quick freeze shelf can become very hot during the defrosting process.
 Therefore, do not touch it!

2.5 Operating the instrument

 Take care when handling microtome knives and disposable blades. The cutting edge is extremely sharp and can cause serious injuries!



We strongly recommend using the safety gloves included with the standard delivery!

- Never leave knives and knife/blade holders with a knife/blade mounted lying around!
- Never place a knife on a table with the cutting edge facing upward!
- Never try to catch a falling knife!
- Always clamp the specimen first, then the knife.
- Before handling the specimen or the knife, changing the specimen or taking a break, lock
 the handwheel and ensure that the knife is covered by the knife guard.
- Extended skin contact with the cold parts of the instrument may cause frostbite!
- To make sure that the condensation water stemming from the defrost cycles drains into the waste container and to avoid the risk of possible contamination, ensure that the tap of the waste container
 (2, Fig. 54) is open whenever operating the cryostat. Only shut the tap when draining the waste container!

2.6 Cleaning and disinfection

We strongly recommend defrosting the CM1860/CM1860UV Cryostat completely for 24 hours, 1-2x annually or as needed.

 It is not necessary to remove the microtome for routine disinfection. The instrument has been designed for UV disinfection. Spray disinfection with Leica Cryofect* is also possible, thanks to the special insulation of the microtome.



Remove section waste after EVERY change of specimen. Do not start disinfection until you have done this! Each new specimen is a potential source of contamination.



The use of flammable sprays within the cryostat chamber is permitted only with the Leica Cryofect tested by us.

At cryochamber temperatures of 0 °C to -35 °C! Spray a **MAXIMUM** of 10 sprays into the middle of the cryostat chamber and close the sliding glass door. After each disinfection, the chamber must be wiped carefully with a cloth and briefly ventilated!

^{*} Cryofect is not sold in all parts of the world

Do not use organic solvents or other aggressive substances for cleaning.
 Use only disinfectants listed in these Instructions for Use, such as Leica Cryofect (alcohol or other common alcohol-based disinfectants).



Please contact Leica Biosystems for further details on adequate disinfection measures.

2.7 Removing the microtome

 The microtome is encapsulated and therefore does not require removal by the user.

2.8 Maintenance

Replacing the fuses

- Turn the instrument off and disconnect the power plug before replacing the fuses!
- Use only the fuse types specified in Chapter 3, "Technical Data". Failure to comply with this can cause severe damage to the instrument and its surroundings!

Replacement of the lamp/UVC lamp



If both disinfection indicator lights are flashing alternately, the UV tube must be replaced!





 Turn the instrument off and disconnect the power plug before replacing the lamps. (For more details, refer to Chapter 9, "Cleaning, Disinfection, Maintenance.)



It is possible to break the UVC lamp during replacement. If this happens, the lamp change must be completed by Technical Service. If any metallic mercury is released, handle it carefully and dispose of it properly.

3. Technical Data

Туре	CM1860/ CM1860 UV				
Test symbol	-	c_CSA_us	-	-	-
Nominal voltage	100 VAC	120-127 VAC	220-230 VAC	220-230 VAC	240 VAC
	±10 %	±10 %	±10 %	±10 %	±10 %
Nominal frequency	50/60 Hz	60 Hz	60 Hz	50 Hz	50 Hz
Power input	1440 VA				
Max. start-up current for 5 sec	2 45 A eff.	45 A eff.	45 A eff.	45 A eff.	45 A eff.
Protective class	1	I	I	I	I
Pollution degree ①	2	2	2	2	2
Overvoltage inst. category	II	II	II	II	П
Heat emission (max.)	1600 J/s				

¹ according to IEC-1010, UL 3101



Operating temperature range (ambient temperature): $18\,^{\circ}$ C to $35\,^{\circ}$ C. All specifications related to temperature of the cooling unit are valid only for an ambient temperature of $22\,^{\circ}$ C and a relative humidity of no more than $60\,\%$!

Refrigeration system	50 Hz/60 Hz
nchigeration system	30 112/00112

Cryochamber

Temperature range $0 \,^{\circ}\text{C}$ to $-35 \,^{\circ}\text{C}$ (+ 3 K/- 3 K)

Cooling time down to -35 $^{\circ}$ C max. 6 hrs., at 22 $^{\circ}$ C ambient temperature

Defrost Automatic hot gas defrost

1 automatic defrost cycle/24 hours, time-controlled (duration 12 min.)

Refrigeration capacity® 690 W Safety factor 3

Refrigerant 300g ±5g refrigerant R 404A * Compressor oil 0.6 I EMKARATE RL-22S, ICI *

Quick-freeze shelf

Maximum temperature $-40 \,^{\circ}\text{C} (+3 \,\text{K/-} 5 \,\text{K})$

Number of freezing stations 8

Defrost Manual hot gas defrost, time-controlled (duration 12 min.)

Peltier element

Max. temperature difference 17K, at - 35 °C chamber temperature Number of freezing stations 2

Number of freezing stations Defrost

In conjunction with the quick-freeze shelf

② according to CECOMAF Liquid temperature 45 °C, evaporation temperature -25 °C



*) Refrigerant and compressor oil must be replaced by qualified, authorized service personnel only.

Microtome

Rotary microtome

Section thickness selection: 1 - 100 µm
Total specimen feed 25 mm
Vertical stroke 59 mm
Maximum specimen size 55 x 55 mm

or 50 x 80 mm

Specimen orientation 8° (x, y, z-axis)

Electric coarse feed

Slow $600 \mu m/s$ Rapid $900 \mu m/s$

Lamp

60 Hz version:

50 Hz version: Osram Dulux S 11 W/21

Color: LUMILUX light white Osram Dulux S 13 W/21

Color: LUMILUX light white

Cryocabinet

Dimensions

Width (w/o handwheel) 600 mm
Width (with handwheel) 730 mm
Depth 730 mm
Height 1140 mm

Weight

(incl. microtome,

without specimen cooling) approx. 135 kg

Storage conditions

Temperature: 5 - 50 °C

Humidity: <80 %, non-condensing

Transport temperature: -40 °C to +55 °C



Observe item 4.1, "Installation site requirements".

All CM1860/CM1860 UV instrument variants need the following secondary fuses:

T0.25A	L250V	Type FST 6.3x32	Brand: SCHURTER
T1.6A	L250V	Type FST 6.3x32	Brand: SCHURTER
T1.0A	L250V	Type FST 6.3x32	Brand: SCHURTER
T6.25A	L250V	Type FST 6.3x32	Brand: SCHURTER
T4A	L250V	Type FST 6.3x32	Brand: SCHURTER
	T0.25A T1.6A T1.0A T6.25A T4A	T1.6A L250V T1.0A L250V T6.25A L250V	T1.6A L250V Type FST 6.3x32 T1.0A L250V Type FST 6.3x32 T6.25A L250V Type FST 6.3x32

4.1 Installation site requirements



Do not operate in rooms with explosion hazard! To ensure proper function of the instrument, it must be set up with a minimum distance on all sides from walls and furniture (see "Transport and installation").

The place of installation must meet the following requirements:

- The instrument requires an installation area of approx. 600 x 730 mm.
- Room volume must be at least 8 m³.
- Room temperature consistently 18 °C 35 °C.
- Temperature range during storage: 5 °C 50 °C
- Relative humidity, maximum 60 % (non-condensing),
- Elevation: up to a max. of 2000 m above sea level



Condensation water may form in the instrument if there is an extreme difference in temperature between the storage location and the installation site and if air humidity is high at the same time. If this is the case, wait at least two hours before switching on the instrument.

Failure to comply with this may cause damage to the instrument.

- The instrument is designed for indoor use only.
- The power plug/circuit breaker must be freely and easily accessible. The power socket may not be located further than 2.5 m from the instrument.
- Power supply at a distance no greater than the length of the power cable an extension cable must NOT be used.
- The floor must be largely vibration-free and have sufficient load capacity and rigidity for the weight of the instrument.
- Avoid impacts, direct sunlight, and excessive temperature fluctuations.
- The instrument MUST be connected to a properly grounded socket. Use only the power cable provided, which is intended for the local power supply.
- The chemicals to be used are easily inflammable and hazardous to health. Therefore the
 installation location must be well ventilated, and must contain no sources of ignition of any
 kind.
- Furthermore, this instrument must NOT be operated directly under the outlet of an air-conditioning system, since the increased air circulation accelerates icing of the chamber.
- The installation location must be protected against electrostatic charge.

Noise information: A-weighted noise level: \leq 70 dB (A)



Room temperatures and humidity levels in excess of the recommendations above will affect the cryostat's cooling capacity and the lowest stated temperatures will not be reached.

4.2 Unpacking instructions for the cryostat CM1860/1860 UV





When the instrument is delivered, check the tilt indicators on the packaging.

If the arrowhead is blue, the shipment was transported laying flat, was tilted at too great an angle or fell over during transport.

Note this on the shipping documents and check the shipment for

Note this on the shipping documents and check the shipment for possible damage.



Caution when removing the metal strips! There is a risk of injury when opening them (the strip has sharp edges and is under tension)!

Fig. 4



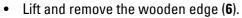
To remove the metal strips (1), you need metal shears and suitable gloves. Stand next to the crate and cut the strips at the location shown (see Fig. 2a "4").

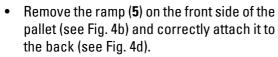


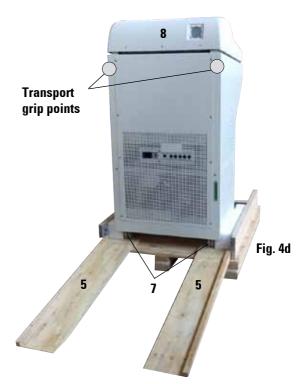
- Lift the outer carton ring (2) upwards and out.
- Remove the foam transport anchors (3), and then pull the dust cover from the instrument.
- Remove all accessories (4).

4. Setting up the Instrument











Do not push the device by its hood (8)! Use the transport grip points (9 ())!

The rollers (7) must run over the ramp (5) at the front and rear. Danger of tipping!

- · Carefully roll the instrument backwards over the ramp from the pallet.
- Push the instrument to the installation location on the castors (7).



The instrument must be transported in an upright position.

Transport to the site

- First, check if the location meets the conditions specified in 'Installation site requirements'.
- Transport the instrument to the desired location.
- · Observe the following:



The instrument must be transported in an upright position or slightly tilted (max. 30°)!



When tilting the instrument 2 people must counterbalance from the front side to prevent the instrument from falling down and causing severe injury, as well as possible severe damage to the instrument and the surrounding area!



4. Setting up the instrument

4.3 Standard delivery—packing list

Basic instrument in the respective voltage variant:

1	knob with marking, antibacterial	14 0491	42558
1	Set of specimen discs	14 0470	43550
	- 4 specimen discs, 25 mm	14 0416	19275
	- 4 specimen discs, 30 mm		
1	section waste tray	14 0471	30787
1	storage shelf, right	14 0491	46599
1	storage shelf, left	14 0491	46598
1	brush shelf		
1	cover for freeze shelf	14 0491	46873
1	toolset:	14 0436	43463
	- 1 brush, fine	14 0183	28642
	- 1 "Leica" brush with magnet	14 0183	40426
	- 1 Allen key, no. 1.5		
	- 1 Allen key, no. 2.5	14 0222	04137
	- 1 Allen key, no. 3.0	14 0222	04138
	- 1 Allen key, no. 4.0	14 0222	04139
	- 1 Allen key with spherical head, no. 4.0	14 0222	32131
	- 1 Allen key, no. 5	14 0222	04140
	- 1 Allen key with handle, no. 5	14 0194	04760
	- 1 Allen key, no. 6	14 0022	04141
	- 1 Single-head wrench, No. 13	14 0330	33149
	- 1 Single-head wrench, No. 16	14 0330	18595
1	bottle of cryostat oil, type 407, 50 ml	14 0336	06098
1	bottle of cryocompound		
	(Jung tissue freezing medium) 125 ml	.14 0201	08926
1	cut-resistant safety gloves, size M	14 0340	29011
1	Instructions for Use, German/English (+ language CD)	14 0491	80001

Compare the delivered components with the parts list and your order. Should you find any discrepancies, please contact your Leica sales office without delay.



A choice of different knife holders is available for the CM1860/ ${\rm CM1860UV}.$

4.4 Assembling the handwheel



The handwheel and all parts for installation are included in the carton with the accessories. The handwheel can be dismounted for transporting (e.g. narrow doors).



Fig. 10

To install the handwheel, follow these steps:

- Insert the pin (1) of the handwheel shaft into the hole (2) of the handwheel.
- Attach the spring washer (3) to the screw (4) as shown in Fig. 10.
- Tighten the screw (4) using an Allen key (6 mm).
- Attach the cover disk (self-adhesive not pictured).

To dismount, proceed in reverse order.



Rotate the handwheel only if the refrigeration system is on and the cryochamber is cold.

5.1 Electrical connection



During the start-up of the refrigeration system, the minimum nominal voltage must not drop below the values specified in Chapter 3, "Technical Data".

The compressor draws a starting current between 45 and 50 A.

Therefore, the electric circuit at the installation site must be inspected by an electrical engineer to ensure that it meets the requirements for a smooth operation of the instrument. A stable power supply in accordance with the instrument's specifications is essential to its proper functioning.

Failure to comply with the above can damage the instrument.

- Have separate fuse protection provided for the electric circuit.
- Do not connect any other appliances to this electric circuit.
- The instrument is de-energized when the power cable is separated from the power supply (power supply circuit breaker).

5.2 Prior to operation

• Check that the power supply voltage and power frequency in your laboratory match the values on the nameplate of the instrument (Fig. 1).



Use only the power cable provided by Leica. Failure to observe this causes an imminently hazardous situation that may result in death or serious injury to the operator or other personnel.

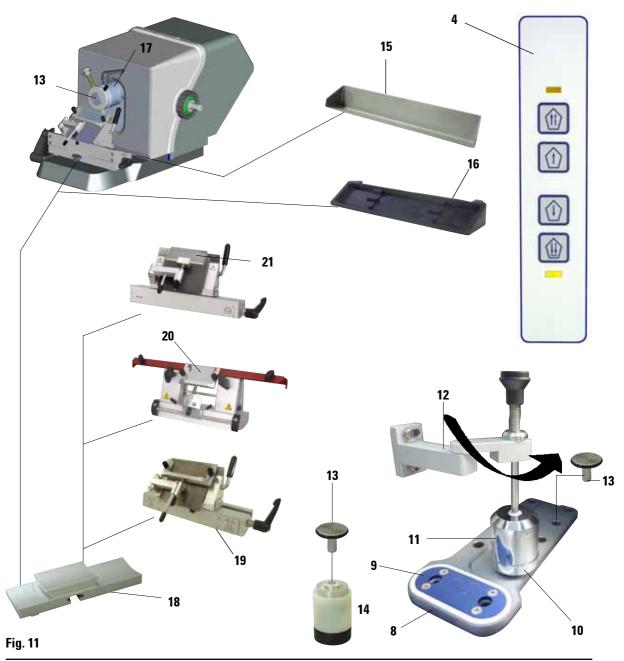
 Check that the drain plug is tight (in the drain hole below the left storage shelf), and press it tightly into the drain if necessary.



The drain plug must be tight in the instrument while work is being carried out in the cryostat. Remove the plug only during the defrost procedure.

- Place the storage shelves in the cryochamber.
- Place the section waste tray and brush shelf in the cryochamber.
- Place the knife holder base on the microtome base plate.
- Set and clamp the blade/knife and knife holders in place (see Chap. 7.3).
- If using a knife, remove it from the box to precool in the cryochamber. Open the knife box with the knife and place it in the cryochamber for precooling.
- Place all tools needed for specimen preparation in the cryochamber.
- Close the sliding window.
- Plug the power plug into the power socket.

5.3 Overview of the Leica CM1860/CM1860 UV



5. Instrument Setup



5.4 Power switch and circuit breaker



The switch must be in the top position (switch position 1) for switching on the circuit breaker.

The switch must be in the bottom position (switch position 0) for switching off the circuit breaker.

Fig. 13
Automatic cutout

5.5 Switching on the instrument



After transporting, wait at least 4 hours before turning the instrument on. This waiting period is necessary to allow the compressor oil, which may have been displaced during transport, to return to its original position. Failure to comply with this may cause damage to the instrument.



The software version "0.40" (shown here only as an example) is displayed in the 4-digit display for the time. This display disappears after 2 seconds and switches to the display of the instrument type "1860". The standard time is then displayed.

- Switch the instrument on at the circuit breaker. The instrument is now initialized.
- The instrument is factory-configured as follows:

Time: 00:00
Defrost time: 23:45
Chamber cooling: On

(Indication of the temperature)

Peltier element: Off

(Display "PE")

• Program the desired values as described under 6.2.1 to 6.2.3.



In normal operation, a pressure compensation shortly before the compressor starts up might lead to a hissing sound.

6.1 Control panel 1



Fig. 14

Function keys



Lamp button

ON/OFF switch for cryochamber illumination.



Manual defrost button

To activate and deactivate manual defrosting.



Key button

To lock and unlock the control panel to protect the entered parameters from unintended modifications. To lock or unlock, hold down for approximately 5 seconds.



Before UV disinfection, move the anti-roll guide to the side to allow complete disinfection.

Opening the sliding window interrupts the disinfection cycle. Press the UVC key to resume the disinfection cycle!



Disinfection

Duration - 30 min

Duration - 180 min



UVC key - to activate / deactivate the disinfection cycle and/or to acknowledge interruption of a disinfection cycle.



For information on the service life of and on replacing the UVC lamp, refer to Chapter 9.3.3.

To start disinfection, the sliding window must be completely closed.

Press the UVC button once briefly, 30 min. mode

Press the UVC button once for a longer time (approx. 4 sec) to start the 180 min. mode

Both LEDs on the display for short and long disinfection indicate the status of the disinfection process:

Status	LED Short disinfection	LED Long disinfection	UVC lamp
Ready ¹	OFF	OFF	OFF
Service life expired ²	alternating flashing		OFF
Short disinfection active	ON	OFF	ON
Long disinfection active	0FF	ON	ON
Short disinfection interrupted ³	Flashing	OFF	OFF
long disinfection interrupted ⁴	OFF	Flashing	OFF

¹ Status: Ready

Both LEDs and the UVC lamp are off.

- Start short disinfection: Press the UVC button briefly (<1 second). The LED for the short disinfection
 cycle will illuminate.
- Start long disinfection: Press the UVC button for longer (>1 second). The LED for the short disinfection goes out and the LED for the long disinfection goes on.

² Status: Service life of the UVC lamp expired

The LEDs for the short and long disinfection modes illuminate alternately, and the UVC lamp is off. This status is displayed as soon as the service life of the UVC lamp (9000 hours) has been reached. For replacing the UVC lamp, refer to Chapter 9.3.3.

6. Operating the Instrument



After replacing the UVC lamp, its service life counter must be reset. To do so, press and hold the UVC button for longer than 30 seconds. The counter has been reset successfully if the LED for the long disinfection goes out while the button is being pressed. Once the UVC button is released, the disinfection status goes into ready mode.

³ Status: Short disinfection interrupted

The LED for short disinfection flashes, and the UVC lamp is off. The operator must press the UVC button to confirm. The disinfection system goes into ready mode, or, if the service life of the UVC lamp has expired, both LEDs flash in alternately.

⁴ Status: Long disinfection interrupted

The LED for long disinfection flashes, and the UVC lamp is off. The operator must press the UVC button to confirm. The disinfection system goes into ready mode, or, if the service life of the UVC lamp has expired, both LEDs flash alternately.



The disinfection mode is interrupted as soon as the sliding window of the cryostat is open or the operator presses the UVC button. In the case of a power failure, when the instrument restarts it checks whether it was running a disinfection cycle, and if so which one. The LED of the determined disinfection cycle (short or long disinfection) flashes.

Programming the desired values 6.2

6.2.1 Setting the time



Use the function button labeled with a clock symbol to set the time of day. To do so, set the current time using the + and - keys.

Pushing the + or - button for more than 1 sec. increases or decreases the time continuously (auto-repeat function).

6.2.2 Setting the automatic defrost time (cryochamber)



Fig. 16

The automatic defrost cycle takes place once every 24 hours.

Briefly touch the + or - button to display the beginning of the defrost cycle which has currently been set. The LED's between the indication of hours and minutes will flash.

To change the beginning of the defrost cycle in 15-minute increments, press the + or - button. The defrost time is 12 minutes.

6.2.3 Selecting the cryochamber temperature



Fig. 17

The temperature of the cryochamber is set and indicated on the panel marked with the cryostat symbol.

The actual temperature is the standard indication. Briefly touching the + or the — button displays the target temperature.

You can now set the desired value using these buttons. Pressing the + or button for more than 1 sec. increases or decreases the chamber temperature continuously.

The actual value will be indicated 5 seconds after finishing the programming.

6.2.4 Activation of the Peltier element (option)



Fig. 18

The Peltier element provides additional cooling for the freezing stations. Upon activation of the Peltier element, the compressor of the cooling system is started after 40 seconds to reinforce the thermal conductivity effect.

Display "PE" (= Peltier element).

Activate it by pressing the *** key.

If the Peltier element has been activated (Fig. 18a), a blue LED in the Peltier symbol lights up. After the activation, the display switches to the number "10" (the Peltier element now cools down for 10 minutes). The remaining cooling time is always shown in the display.

The Peltier element turns off automatically after 10 minutes.

When only 4 minutes are remaining, a dot appears after the number 4 ("4."). From this time on, it is safe to shut off the Peltier element early.

To shut it off early, press the *** key again.

After the shutoff, the display returns to "PE".



Fig. 18a

6.2.5 Manual defrosting of the quick freeze shelf





Fig. 19



The quick freeze shelf may become hot during defrosting! Therefore, do not touch it!

You can activate the manual defrost of the quick freeze shelf by pressing the key (continuous tone sounds), followed immediately by the key (continuous tone stops). The display flashes during the defrosting (duration 12 min.).

To shut off the manual defrost early, press the key, followed immediately by the key. The quick freeze shelf can become hot during the defrosting process. The defrost time is 12 minutes.



The quick freeze shelf and cryochamber cannot be defrosted at the same time.

6.2.6 Manual defrosting of the cryochamber





You can activate manual defrosting of the cryochamber by pressing the key (continuous tone sounds), followed immediately by the + key in the cryochamber temperature selection field (continuous tone stops). The display flashes during the defrosting (duration 12 min.).

To shut off the manual defrost early, press the key, followed immediately by the button in the cryochamber temperature selection field.

Fig. 20

6.2.7 Setting the section thickness



Fig. 21

Adjust the section thickness using the $\boxed{+}$ and $\boxed{-}$ pushbuttons in the top control panel field (see Fig. 21) in a range from 1 - 100 μ m:

From 1 - $5 \mu m$ in $0.5 \mu m$ increments

From 5 - 20 μ m in 1 μ m increments

From 20 - 60 µm in 5 µm increments

From 60 - 100 µm in 10 µm increments

The preselected section thickness appears in the display above the keys.

- Start trimming at approx. 20 μm.
- Decrease the section thickness continually down to the appropriate value.
 After changing the section thicknesses, discard the first two or three sections.
- For sectioning, turn the handwheel at a consistent speed.

6.3 Retraction (factory default setting: disabled)

Enabling retraction:

When switching on the instrument (in the initialization phase), you have to press the + key (Fig. 21). If the handwheel is at the 6 o'clock position or the specimen head is "down", the yellow LED next to **RETRACT** is illuminated. When the instrument is switched off, this status is stored.

Disabling retraction:

When switching on the instrument (in the initialization phase), you have to press the + key. If the handwheel is at the 6 o'clock position or the specimen head is "down", the yellow LED next to **RETRACT** is not illuminated and remains off even during sectioning.

6.4 Display lock





Fig. 22

After the display is locked using the key button (press and hold for 5 sec.), set values can **no longer** be changed.

Only the UV disinfection can still be enabled/disabled by pressing the UV key. Push the key button once more for 5 seconds to unlock the display.

When the display is locked, the LEDs between the hour and minute indication on the time panel are turned off.

Control panel 2 - Electric coarse feed 6.5



Move the specimen away from the knife

Starts fast retraction to the rear end position.

The LED (1) flashes while the specimen head is in motion.

The LED (1) lights up when the rear end position has been reached.

- The return movement can be stopped by pressing one of the coarse feed buttons.
- Starts slow retraction to the rear end position.

The motion continues as long as the button is held.



slow



slow



Fig. 23

2

Advancing the specimen toward the knife

Starts fast or slow feed toward the knife. The LED (2) starts flashing as long as the specimen head is in motion.

The LED (2) lights up when the forward end position has been reached.



This is a safety feature designed to protect both the specimen and the knife.

To feed the specimen, press or hold the appropriate SLOW or RAPID button.

7.1 Specimen freezing

 Select the sectioning temperature (chamber temperature) according to the tissue type to be sectioned (see "7.4 Temperature selection chart").

7.1.1 Quick freeze shelf

The cryochamber is equipped with a quick freeze shelf (5, Fig. 24) on which up to 10 specimen stages with specimens can be attached.

The temperature of the quick freeze shelf is always lower than the cryochamber temperature.



Fig. 24

- Cut the specimen to size.
- If necessary, enable the Peltier element (4) it takes approx. 60 seconds until the maximum cooling capacity is available.
- Apply a sufficient amount of cryocompound to a specimen disc to orient and mount the specimen.
- Place the specimen disc in one of the holes of the quick freeze shelf and freeze the specimen at a low temperature.
- Once the specimen is frozen, insert the specimen disc (Fig. 25) with the specimen into the specimen head (Fig. 25) and start sectioning.

7.1.2 Stationary heat extractor (optional)

- Fasten the support (1, Fig. 24) of the heat extractor (2, Fig. 24) to the left side panel of the cryochamber using two screws in the provided holes. Insert the heat extractor.
- Lower the heat extractor cylinder on the specimen surface. After approximately 30 seconds, the specimen should be completely frozen.
- After the specimen is frozen, move the heat extractor into resting position at the parking station (3, Fig. 24).

7.2 Specimen discs

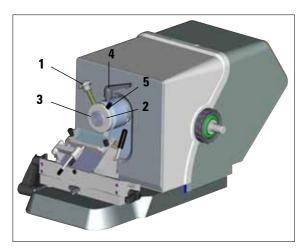


Fig. 25

7.2.1 Inserting the specimen discs into the specimen head

- Lock the handle of the handwheel in the upper position.
- If the knife holder and a knife are in place, cover the knife edge with the knife guard. If using a CE disposable blade holder, the antiroll guide should be lowered to cover the blade edge.
- Loosen the clamping screw (1, Fig. 25) on the specimen head.
- Insert the shaft of the specimen disc (3) in the location hole (2) of the specimen head.



Make sure that the shaft of the specimen disc is fully inserted. The entire surface of the rear side of the specimen disc must be free of residue.

• Tighten screw (1).



- Loosen the clamping lever (4).
- Orient the specimen on the ball shell using the lever (5).
- Tighten the clamping lever (4).



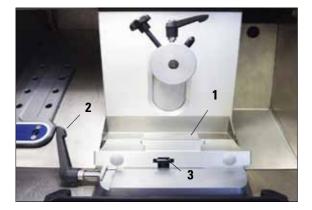
The specimen discs are to be handled by their side O-rings to protect against frostbite!

7.3 Sectioning

7.3.1 Blade holder CE



If an additional blade holder instruction manual has been provided, read and follow the instructions and safety information carefully!



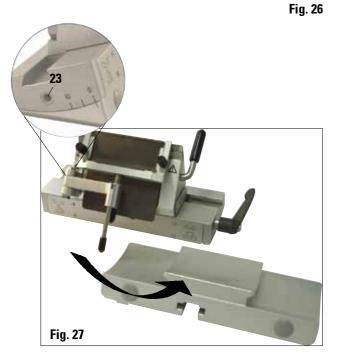
Inserting the base

 To insert the blade/knife holder base (1), move the lever (2) to the back. Slide the base onto the T-piece (3) of the baseplate.



When inserting the blade/knife holder base, you have to overcome the resistance of a spring (located in the foot of the knife holder) by applying a little pressure towards the left.

2. To clamp the blade/knife holder base, turn the lever (2) to the front.



Inserting the blade holder CE

3. Push the upper part of the blade holder onto the base. Clamp the blade holder into place using an Allen key (No. 4) on the left side (23).

7. Daily Use of the Instrument

Inserting the blades into blade holder CE



Caution!

Microtome blades are extremely sharp!



The blade holder CE can be used either for the low-profile, narrow-band and high-profile, broad-band blades.

Inserting the high-profile blades

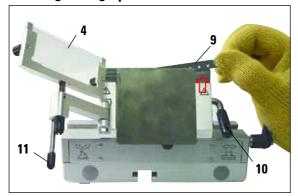
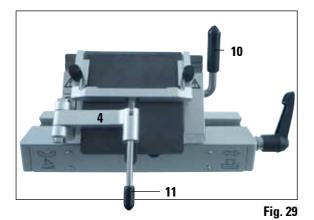


Fig. 28



Inserting low-profile blades



Fig. 30



The safety gloves included in the standard scope of delivery must be worn when inserting the blade!

- Fold the anti-roll guide system (4) to the left—while doing so, hold the lever (11) (not the adjusting screw of the anti-roll guide) so that the height of the anti-roll guide remains unchanged.
- 2. Open the clamping lever (10) by turning it counterclockwise (Fig. 28).
- 3. Carefully insert the blade (9) from above or from the side between the pressure plate and the blade rest. Make sure that the blade is inserted so that it is centered and lies evenly along the ledge (see red arrow in Fig. 28).
- 4. Clamp the clamping lever (10) by turning it clockwise (see Fig. 29).
- 5. Fold the anti-roll guide system (4) back to the right (toward the blade) using the lever (11).



The anti-roll guide system functions as a knife guard here!

 When using low-profile blades, the red guide (blade rest) must first be placed in the blade holder, and then the blade.

Inserting low-profile blades (continued)

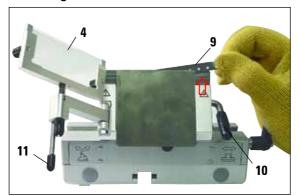


Fig. 31

Two magnets are attached to the rear side of the guide. These point away from the operator after the guide has been inserted (towards the rear pressure plate).

Then insert the blade as described (for high-profile blade).

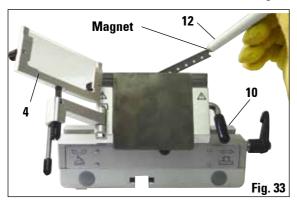
Removing the blades



- Fold the anti-roll guide system (4) to the left

 while doing so, hold the lever (11) (not the
 adjusting screw of the anti-roll guide) so that
 the height of the anti-roll guide remains un changed.
- 2. Open the clamping lever (10) by turning it counterclockwise (Fig. 32).
- **3**. Carefully lift out the blade (9). Continue with instruction 5.

Fig. 32



4. Another option for removing the blade is to use the brush with magnet (**12**).

To do so, fold the clamping lever (10) downwards counterclockwise (Fig. 33). Fold the anti-roll guide system (4) to the left.

Guide the brush with magnet to the blade and lift it upwards and out.

7. Daily Use of the Instrument





The safety gloves included in the standard scope of delivery must be worn when disposing of the blade!

Once the blade has been removed from the blade holder, it is disposed of into the dispenser container (storage compartment in bottom, Fig. 34).

Fig. 34

Lateral movement

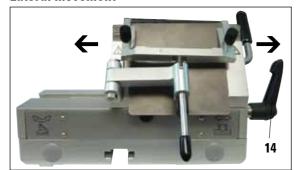


Fig. 35

It is only necessary to clamp a disposable blade into the CE holder one time. When a portion of the blade becomes dull or nicked, the upper portion of the blade holder (shown on its base in Fig. 35) will move from side to side to allow the use of the entire blade edge. When moved, the alignment of block to blade will be very close to the previous position since the clamping pressure on the blade remains the same.

To reposition the blade holder, follow these steps:

- Turn the clamping lever (14) to the rear (clockwise) to release, and then shift the blade holder sideways to the desired position.
- 2. To tighten, turn the clamping lever (14) to the front (counterclockwise).

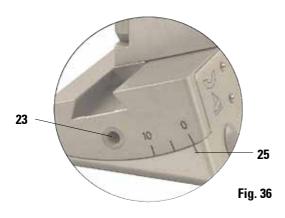
Clearance angle adjustment



The harder the specimen is, the larger the clearance angle must be. However: The larger the clearance angle is, the larger the section buckling becomes.

If the clearance angle is too small, sections that are too thick or thin can result. Also consider that the position of the knife edge relative to the specimen can change when the clearance angle is adjusted. For this reason, always place the specimen above the knife when adjusting the clearance angle. Otherwise, the specimen could hit the knife while being raised.

A clearance angle that is too steep or too shallow may not lead to optimum sectioning results
and can possibly damage the specimen. We generally recommend using higher clearance
angles for harder specimens and relatively lower clearance angles for softer ones.



Adjust the clearance angle with every new tissue type!

- 1. The clearance angle scale is located on the left side of the blade holder.
- Release the blade holder by turning the No. 4 Allen screw (23) counterclockwise. Select a clearance angle of 0°. To do so, align the number 0 to the index mark (25) and tighten the Allen screw (23). If the sectioning results are not satisfactory, increase the clearance angle in 1° increments until you achieve optimum results.



Settings of 1° - 2° (blade holder CE) are well suited for most applications.

Adjusting the anti-roll guide system

You can adjust the height of the anti-roll guide system using the knurled nut (8):

- If you turn the nut counterclockwise, the anti-roll guide system moves toward the blade.
- If you turn the nut clockwise, the anti-roll guide system moves away from the blade.

If the anti-roll guide system is in the wrong position relative to the blade, the following problems will result:



Fig. I: The section rolls over the glass insert of the anti-roll guide system.

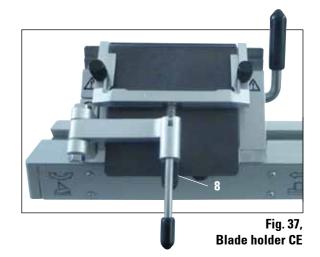
Error: Glass insert not high enough.

Remedy: Turn the knurled nut counterclockwise until the section is pushed between the blade and antiroll guide as shown in Fig. III.

Fig. II: Section is compressed and block contacts and moves the glass insert during the upstroke.

Error: Anti-roll guide system is set too high.

Remedy: Turn the knurled nut clockwise until the section is pushed between the blade and anti-roll guide as shown in **Fig. III**.

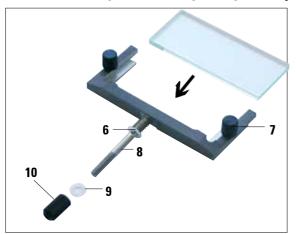






We ALWAYS recommend pre-adjusting the anti-roll guide system at a high section thickness (e.g. 10 μ m). — Start from there and work your way down to the desired section thickness in small increments, readjusting the anti-roll guide system at each increment using the knurled nut.

Assembling the anti-roll guide system/replacing the anti-roll plate



the pin rests in the notch.

3. Push the white plastic plate (9) from below onto the shaft (8).

Insert the glass into the interchangeable frame and tighten it evenly using the knurled screws (7).
 Insert the shaft (8) of the metal frame for exchangeable glass inserts from above into the bore of the swinging arm (6) in such a way that

4. Screw the knurled nut (10) from below onto the shaft (8).



All 4 lengths of the anti-roll guide glass can be used.

Fig. 38

7.3.2 Blade holder CE-TC

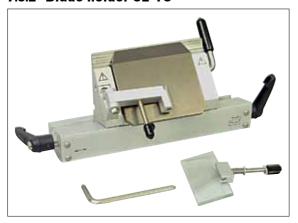


Fig. 39

The blade holder CE-TC is designed only for disposable carbide metal blades (tungsten carbide - TC65).

The handling of this holder is identical to that of the blade holder CE.

7. Daily Use of the Instrument

7.3.3 Knife holder CN

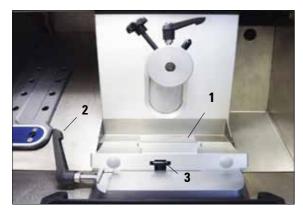


Fig. 40

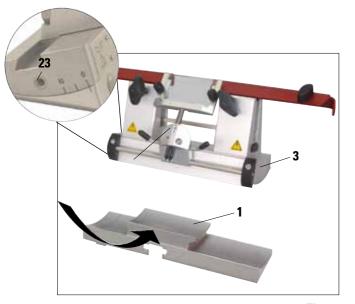


Fig. 41

Inserting the base

 To insert the blade/knife holder base (1), move the lever (2) to the back. Slide the base onto the T-piece (3) of the baseplate.



When inserting the blade/knife holder base, you have to overcome the resistance of a spring (located in the foot of the knife holder) by applying a little pressure towards the left.

2. To clamp the blade/knife holder base, turn the lever (2) to the front.

Inserting knife holder CN

3. Push the upper part of the knife holder (3) onto the base (1). Clamp the knife holder into place using an Allen key (No. 4) on the left side (23).



Clearance angle settings of 4° - 6° (knife holder CN and blade holder CE-TC) are well suited for most applications.

Inserting/removing the knife

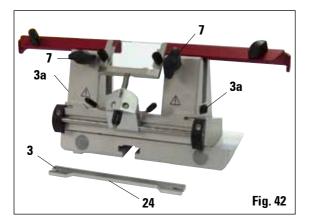


The height of resharpened knives must be adjusted using knurled screws (3a) (approx. 1 mm under the edge of the clamping jaw).

Make sure that the knife is parallel from one end to the other.

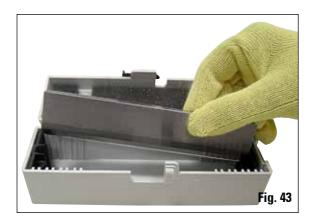


The safety gloves included in the standard scope of delivery must be worn when inserting and removing the knife!





After removing the knife from the knife holder, place it safely into the knife case. NEVER place it onto the work surface next to the instrument!

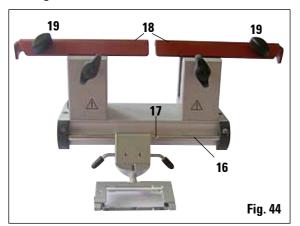


- Insert the long knife support (3) on the side above the knurled screw (3a) so that the recess (24) faces the user – turn the height adjustment knurled screws until the lower stop is reached.
- The knife can now be inserted from the side and its height adjusted via the knurled screws (3a). The upper edge of the rear clamping jaw serves as an index point for the correct knife height. The height of the knife blade should match that of the rear clamping jaws. This makes it possible to accurately adjust even greatly worn knives up to a height of 25 mm.
- Once the correct height is obtained, tighten the wingscrews (7) in alternating order until both are tight.
- To remove the knife, follow these instructions in opposite order.

Open the wingscrews by turning them counterclockwise and then pull the knife out to the side.

7. Daily Use of the Instrument

Knife guard/lateral movement for knife holder CN



The knife guard (18) is fixed and integrated into the clamping jaws. The knife guard has handles (19) that allow it to be moved. The knife guard is suitable for knife lengths up to 16 cm. Always cover exposed parts of the knife blade after sectioning. The anti-roll guide system can be moved sideways (only for the 84 mm variant). To better find the mid position, a groove (17) is provided in the shaft (16), in which the anti-roll guide sits.

- Pressure plate permits the full length of the knife to be utilized.
- The CN knife holder can be used to hold tungsten-carbide tipped or steel knives and disposable blade rails.

Clearance angle setting

The clearance angle adjustment for the knife holder CN is carried out exactly as described for the blade holder CE.

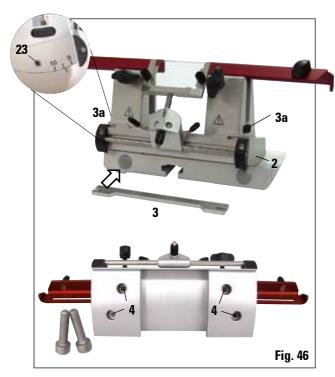
Adjusting the anti-roll guide

The anti-roll guide adjustment for the knife holder CN is carried out exactly as described for the blade holder CE. For lateral adjustments, refer to the section on lateral movement (on this page above).

Knife holder CN with anti-roll guide – Moving the clamping chucks



If large specimen discs are to be used in the specimen holder (e.g. $50 \times 80 \text{ mm}$), it is possible to shift the clamping jaws laterally.



The clamping jaws are factory-installed in the knife holder with a clearance of 64 mm. If necessary, both clamping jaws can be offset with a clearance of 84 mm.

Perform the following steps:

- Use a 4 mm Allen key to loosen the screw over the clearance angle adjustment (23) and remove the segment arc (2) from the blade/ knife holder base.
- Use a 4 mm Allen key to loosen the screws (4) on the underside of the segment arc.



Never work with only one clamping jaw, as this does not ensure the stability required for the sectioning process. Also, a long knife will not be sufficiently protected by the knife guard in this instance.

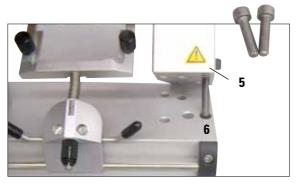


Fig. 47

 Lift off the clamping jaw (5) on the right side (caution: washers) and feed it into the adjacent hole (6). Tighten the screws from the underside of the segment arc. Repeat on the left side.
 Now use the longer knife back rest provided.

7.3.4 Knife holder CN-Z with anti-roll guide

This is used for holding conventional and carbide-metal knives up to 16 cm in length. The pressure plate ensures extreme stability and the full utilization of the knife blade.

The height of the knife edge is adjustable, and the clearance angle adjustment must be adjusted separately (see Knife holder CN).

The knife guard (18) is moveable.

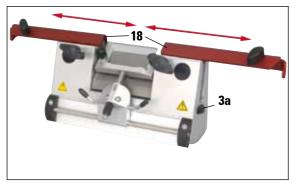


Fig. 48



The height of resharpened knives must be adjusted using knurled screws (3a) (approx. 1 mm under the edge of the clamping jaw).

Make sure that the knife is parallel from one end to the other.

- For this knife holder, the clamping jaws are not offset.
- Pressure plate permits the full length of the knife to be utilized.
- The CN knife holder can be used to hold tungsten-carbide tipped or steel knives.

7.3.5 Cleaning the blade and knife holders



For daily cleaning, remove the section waste from the knife holder using a dry brush. Use a cold brush or section waste will thaw and stick to the blade or knife holder.



Mechanical damage to the pressure plate will strongly affect the sectioning quality. For this reason it is important to ensure that it is not damaged during sectioning, cleaning or other functions.

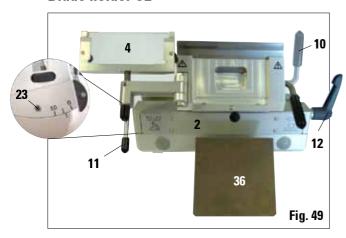


For disinfection, commercially available, mild cleaners and disinfectants can be used – we recommend Leica Cryofect (not available in the USA).

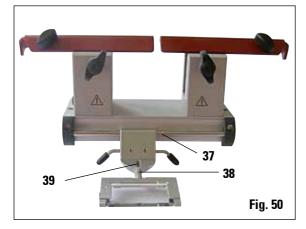
Wear gloves during cleaning to prevent frostbite.

7. Daily Use of the Instrument

Blade holder CE



Knife holder CN



Disinfection

Place all of the removed parts into a disinfecting solution for the time recommended by the manufacturer. Dry each part thoroughly before reassembly and insertion into the cryostat.

Occasionally oil the moving parts such as the axes (37) and (38) and the slot (39) with a drop of cryostat oil (Type 407).

- To release the segment arc (2) from the base, use a 4 mm Allen key to loosen the screw over the clearance angle adjustment (23) and remove the segment arc (2) from the blade/ knife holder base.
- 2. Fold the anti-roll guide system (4) to the left while holding the lever (11).
- 3. Open the clamping lever (10, Fig. 49) by turning it counterclockwise and pull it out.
- 4. Afterwards, the pressure plate (**36**) can be removed for cleaning (with alcohol).
- Open the clamping lever of the blade holder (12, Fig. 49) by turning it counterclockwise and pull it out. The blade holder can now be moved sideways and can be taken out of the segment arc.



If several blade/knife holders are cleaned at the same time, the parts must NOT be mixed up! Failure to adhere to this may result in sectioning problems!



Also observe the warning message on Page 13!

7. Daily Use of the Instrument

Trimming the specimen



Take care when handling microtome knives and disposable blades. The cutting edge is extremely sharp and can cause serious injuries! For that reason, always wear the cutresistant safety gloves included in the standard scope of delivery!



Always clamp the specimen block BEFORE clamping the knife or the blade. Lock the handwheel and cover the knife edge with the knife guard prior to any manipulation of knife/blade or specimen, prior to changing the specimen block and during all work breaks!

- Insert the precooled knife/blade in the knife/blade holder.
- Adjust the appropriate clearance angle on the knife holder. Settings from 4° 6° (knife holder CN and blade holder CE-TC) or 1° 2° (blade holder CE) are suitable for most applications.
- Align specimen with the knife holder/knife or blade holder/blade using the x-y orientation as described on page 36.
- Remove the knife guard (knife holder CN) or fold the glass anti-roll guide (knife holder CE, CE-TC) over to the side.
- Unlock the handwheel.
- To trim the specimen, move the specimen towards the knife using the coarse feed buttons. Trim the specimen down to the required sectioning plane by turning the handwheel. (see 6.2.7 "Setting the section thickness")
- Position the anti-roll guide on the knife and align with the cutting edge.
 Adjust the anti-roll plate if necessary (see "Clearance angle adjustment").

7.3.6 Installing the shelf (movable) (optional)



Attach the rod for the shelf to the inner front side of the cryostat housing using the provided screws (1) and the No. 3 Allen key, then attach the caps (3). (The rear side of the movable shelf has white plastic screws (2) that prevent the interior of the chamber from being scratched.) Now hook the movable shelf into the guide rod.

7.4 Temperature selection chart (in minus °C)

Tissue type	-10°C – -15°C	-15°C – -25°C	-25 °C — -35 °C
Adrenal	*	*	
Bone marrow		*	
Brain	*	*	
Bladder		*	
Breast - fatty			*
Breast - little fat		*	
Cartilage	*	*	
Cervical		*	
Fatty			*
Heart and vascular		*	
Intestinal		*	
Kidney		*	
Laryngeal		*	
Lip		*	
Liver		*	
Lung			*
Lymphoid		*	
Muscle		*	
Nose		*	
Pancreatic		*	
Prostate		*	
Ovarian		*	
Rectal		*	
Skin with fat			*
Skin without fat		*	
Spleen or bloody tissue		*	
Testicular	*	*	
Thyroid		*	
Tongue		*	
Uterus curettage	*		

The temperature values given in this table are based on experience, however, these are only approximate values, as any tissue may require particular adjustments.

7.5 Defrosting

Defrosting the cryochamber actually means defrosting the evaporator to prevent excessive frost buildup. The evaporator is flushed with hot gas during defrosting. The cryochamber virtually is frostfree and does not need to be defrosted.

The condensation water that is produced during defrosting is collected in a container, which is located at the front of the cryostat cabinet.



To make sure that the condensation water stemming from the defrost cycles drains into the waste container and to avoid the risk of possible contamination, ensure that the tap of the waste container (2, Fig. 54) is open whenever operating the cryostat.

Only shut the tap when draining the waste container!



The quick freeze shelf is cooled during the automatic cryochamber defrosting. However, the Peltier element is turned off.

The maximum duration of a defrost cycle is 12 minutes. Defrosting is terminated automatically once the cryochamber has reached a temperature of -5°C. Cooling turns back on automatically.

7.5.1 Automatic defrosting of the cryochamber

An automatic defrost cycle takes place once in 24 hours.

You can program the time of the automatic defrost cycle on control panel field 1 (see item 6.2.2).

7.5.2 Manual defrosting of the cryochamber



To avoid an unintended defrosting, the activation of a manual defrost cycle is confirmed by an audible signal. Cooling turns back on automatically.

In addition to the programmed automatic defrost, a manual defrost for the cryochamber can also be activated (see item 6.2.6).

7.5.3 Manual defrosting of the quick freeze shelf



The quick freeze shelf may become hot during defrosting! Therefore, do not touch it!

In case of heavy frost formation on the quick freeze shelf, which can occur particularly after the spray disinfection, a defrost can be activated manually (see point 6.2.5). which can be terminated when required.



Be sure to place the corresponding cover on the quick freeze shelf to prevent frost. Always cover the quick freeze shelf during breaks and overnight.

7.6 Terminating work

7.6.1 Terminating daily work

- Lock the handwheel.
- Take the knife out of the knife holder and put it back into the knife box in the cryochamber.
- Remove frozen section waste with a cold brush.
- Empty the section waste tray.
- · Clean the storage shelves and brush shelf.
- Remove all tools and supplies that would prevent the UV light from reaching the chamber walls and shelves.



To clean and disinfect the interior of the instrument, use only alcohol-based disinfectants. On the exterior, common detergents and alcohol-based disinfectants can be used.

All components removed from the cold environment will collect condensation. Therefore, dry them thoroughly before placing them back into the cryochamber.

- · Remove all the specimens from the cryostat.
- Attach the cover for the freeze shelf (standard scope of delivery).
- · Remove the plug and close the sliding window.
- Switch off the chamber illumination and start the UV disinfection.
- · Lock control panel field 1 (Fig. 14) using the key button.
- Do NOT switch off the instrument using the circuit breaker, as otherwise no cooling takes place.

7.6.2 Shutdown for a longer period



If you do not intend to use the instrument for several weeks you may turn it off.

Please note, however, that it may take up to several hours to cool the cryochamber down to very low temperatures after turning the instrument on again.

After turning off, the instrument should be cleaned and disinfected thoroughly (see chapter "9. Cleaning, Disinfection, Maintenance").

- Follow the steps in 7.6.1 until UV disinfection is complete.
- Switch the instrument off at the circuit breaker and unplug the power plug.
- Open the sliding window to allow air circulation, allow the chamber to dry and prevent accidental breakage while stored.

Switching off the instrument via the circuit breaker will not affect the programmed values.

Before turning the instrument on again, the cryochamber microtome and all accessory components must be absolutely dry.

8.1 Error messages in the display



Error messages are displayed on the clock panel as follows **EO**: **XX**. The following error messages might occur during operation:

Fig. 52

Error	Cause	Remedy
20	Calibration error; possibly defective controller board.	Switch the instrument off for 10 seconds and then back on. If the error is displayed again: Call service.
21	Clock battery on the controller board empty.	Call service.
23	Cryochamber temperature out of display range of 35°C to -55°C .	Resolve cause.
25	Temperature sensor for cryochamber temperature defective.	Call service.
27	Temperature sensor for defrost limiter defective.	Call service.
28	Both light barriers in the handwheel active.	Call service.
29	Service interval warning after 15960 hours.	Call service.
30	Service interval expired after 17610 hours.	Call service.



If errors occur, they are output as error codes in the real-time display during normal operation with the format EO:XX.

- In normal operation, the error displayed can be acknowledged by pressing any key once. Error codes 21-28 are self-acknowledging if their cause has been eliminated.
- After 15,960 hours, "HELP" is output in the real-time display to indicate the need for service maintenance. The display alternates between "HELP" and the real-time display. The note is acknowledged by pressing a key on the keyboard. The "HELP" display reappears for a longer period.
- After 17,610 hours, "HELP" is output in the real-time display to indicate the need for service maintenance. The display alternates between "HELP" and the real-time display. The note is acknowledged by pressing a key on the keyboard. The "HELP" display reappears for a shorter period.

8. Troubleshooting

8.2 Temperature control button

On the back of the cryostat cabinet there is a temperature control button (1, Fig. 53). If the temperature of the cryochamber exceeds 60 °C the switch is automatically activated and turns the instrument off.

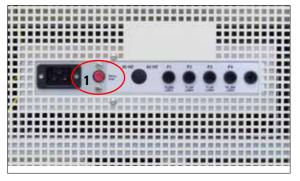


Fig. 53

Possible causes and remedies:

- Temperature of the direct surroundings is constantly higher than 40°C.
- -->Drop the temperature of the direct surroundings.
- When setting up the instrument the minimum distance (see "Transport and installation") to walls and furniture was not kept.
- --> Keep the minimum distance.
- The ventilation slits of the liquefier are dirty.
- --> Clean air inlet openings (see Fig. 56).

After eliminating the possible sources of error, push the temperature control button (1, Fig. 53) to turn the instrument back on and make it ready to operate. If the instrument does not respond, contact service.

8.3 Possible sources of error, causes and remedies

Problem	Cause	Remedy
Frost on chamber walls and microtome	 Cryostat is exposed to air currents (open windows and doors, air conditioning). Sliding window was open and exposed to air currents too long. Frost built up by breathing into the cryochamber. 	 Change place of installation for the cryostat. Close the sliding window correctly. Wear a mask if necessary.
Ice formation on the bottom of the cryochamber	 Condensation water drain obstructed. Drain of the quick freeze shelf defrosting water obstructed. 	 Open the shut-off valve, switch off the instrument and let it thaw and dry. Leveling the instrument.
Sections bunch up at the blade edge	 Specimen not cold enough. Knife and/or anti-roll plate not yet cold enough - the section melts as a result. 	 Select lower temperature. Wait until knife and/or antiroll plate have reached cryochamber temperature.
Sections splinter, cracks in the section,	- Specimen too cold	- Select higher temperature.
Sections not properly flattened	 Static electricity/air currents. Specimen not cold enough. Large area specimen. Anti-roll plate poorly positioned. Anti-roll plate poorly aligned with knife edge. Incorrect clearance angle. Knife/blade blunt or damaged. 	 Eliminate cause. Select lower temperature. Trim the specimen parallel; increase section thickness. Reposition anti-roll plate. Align correctly. Slightly increase knife/blade angle. Use different part of the cutting edge or replace.

8. Troubleshooting

Problem	Cause	Remedy
Sections not properly flattened despite correct temperature and correctly aligned anti-roll plate	Knife/blade and/or anti-roll plate dirty.Edge of anti-roll plate damaged.Blunt knife/blade.	Clean with dry cloth or brush.Replace plate.Use different part of the cutting edge or replace.
Sections curl on the anti-roll plate	 Anti-roll plate does not protrude far enough beyond the cutting edge. 	- Readjust correctly.
Scraping noise during sectioning and specimen return movement	 Anti-roll plate protrudes too far beyond the cutting edge and is scraping against the specimen. 	- Readjust correctly.
Uneven sections	Knife/blade damaged.Edge of anti-roll plate damaged.	Use different part of the cutting edge or replace.Replace plate.
Chatter during sectioning	 Specimen insufficiently frozen onto the specimen disc. Specimen disc not clamped tightly. Specimen holder ball joint not clamped. Knife/blade not clamped tightly enough. Specimen has detached from the disc. Very hard, inhomogeneous specimen. Blunt knife/blade. Knife profile inappropriate for this specimen used. Incorrect clearance angle. 	 Refreeze specimen onto the disc. Check specimen disc clamping. Check ball joint clamping. Check knife/blade clamping. Refreeze specimen onto the disc. Decrease section thickness; reduce specimen surface area if necessary. Use different part of the cutting edge or replace. Use knife with different profile. Slightly decrease clearance angle.

Problem	Cause	Remedy
Condensation on anti-roll plate and knife during cleaning	- Brush, forceps and/or cloth are too warm.	Use components and clean- ing materials only after cool- ing them. Store all tools on storage shelf in the cryochamber.
Anti-roll plate damaged after adjustment	 Plate too high above the cutting edge. The adjustment was car- ried out in the direction of the knife. 	 Replace anti-roll plate; then lift it during adjustment. Handle anti-roll plate more carefully.
Thick-thin sections	 Temperature incorrect for the tissue cut. Knife profile inappropriate for the specimen cut. Ice buildup in the knife back. Handwheel speed not uniform or turned at incorrect speed. Knife not clamped tightly enough. Specimen holder not clamped tightly. Cryocompound applied to cold specimen disc; specimen detached from the disc after freezing. Blunt cutting edge. Incorrect clearance angle. Dried specimen. 	 Select correct temperature. Wait until the correct temperature is reached. Use knife with different profile (c or d). Remove ice. Adapt speed. Check knife/blade clamping. Check specimen clamping. Apply cryocompound on warm disc; mount specimen and freeze. Use different part of the cutting edge or replace. Slightly increase clearance angle. Prepare new specimen.

8. Troubleshooting

Problem	Cause	Remedy
Tissue adheres to the anti-roll guide.	 Dried specimen. Anti-roll plate is too warm or incorrectly positioned. Static charge. Fat on the corner or edge of anti-roll plate. Rust on the knife. 	 Cool down anti-roll plate or reposition correctly. Remove static electricity. Remove fat with alcohol.
Flattened sections curl up when anti-roll plate is folded up.	- Static charge or air current Anti-roll plate too warm.	Remove static electricity. Cool down the anti-roll plate.
Sections tear or separate.	 Temperature too low for the tissue being sectioned. Blunt part, dirt, dust, frost or rust on the knife. Front edge of the anti-roll plate damaged. Hard particles in the tissue. Dirt on back of knife. 	 Increase temperature and wait. Remedy cause (> see individual specific solution). Replace plate. If the application permits, cut deeper during sectioning. Clean.
Inconsistent or insufficient specimen feed not uniform or inaccurate.	- Defective microtome.	- Call technical service.
Specimen disc cannot be removed.	 Moisture on the underside causes the specimen to freeze to the freezing shelf or specimen head. 	- Apply concentrated alcohol to the contact point or heat the specimen head.
Cryostat non-operational.	Mains plug not properly connected.Circuit breaker is switched off.	Check if power plug is properly connected.Move the switch back to the upper click stop.

Problem	Cause	Remedy
No or insufficient refrigeration	Compressor defective.Leak in the cooling system.Inappropriate site conditions.	Call technical service.Call technical service.Check Installation site requirements
	 Ventilation grilles of the condenser dirty. 	(item 4.1) - Clean air inlet opening.
Scraping noise at the slot cover of the microtome	- Friction between the slot cover and the microtome housing.	 Apply cryostat oil to the slit cover and distribute it by moving the handwheel or using a clean cloth.
Both disinfection indicator lights flashing alternately.	 UV radiation provided by UV tube no longer sufficient for disinfec- tion. 	 Replace UV tube following the manufacturer's instruc- tions.

8.3.1 Changing the battery

The battery is exchanged during routine maintenance.

If the customer has decided against a maintenance contract, the battery must be replaced after 7 years at the latest. All settings (time, defrost time, section thickness etc.) are lost when the instrument is switched off.

There is no danger for the instrument. However, the customer must re-enter the setting values after changing the battery.

9.1 Cleaning

- Remove frozen section waste from the cryostat with a cold brush every day.
- Remove the section waste tray for emptying.
- Remove the storage shelves and the brush shelf for cleaning.
- Remove the closed sliding window from the front by gently lifting it (see 9.3.4, "Replacing the fluorescent lamp").



The use of flammable sprays within the cryostat chamber is permitted only with the Leica Cryofect tested by us.

Do not use organic solvents or any other aggressive substances for cleaning and disinfecting!

Wear safety gloves at all times when cleaning, to prevent contact with contaminated substances!

Use only disinfectants listed in these Instructions for Use, such as Leica Cryofect (alcohol or other common alcohol-based disinfectants).



- Drain the cleaning liquid through the hose after the prescribed reagent time is over and collect it in the waste container (1).
- Dispose of liquid according to the waste disposal regulations.
- To remove the waste container (1), shut off the tap (2) and unscrew the lid (3).



The catch tank collects the condensate that accumulates during defrosting. Therefore, check the level regularly and empty the container according to applicable laboratory regulations.

Fig. 54

9.2 Spray disinfection with Leica Cryofect

Basically, we recommend UV disinfection (for application, see page 27).

For easy-to-use spray disinfection we recommend Leica Cryofect. (Leica Cryofect is not available in the USA!)



We recommend wearing the safety gloves provided in the standard scope of delivery when disinfecting!

Exercise caution when cleaning the chamber – the plug in the rear on the left has sharp edges and can lead to cutting injuries.

The cryostat, including all components, has to be disinfected after each daily use.



Comply with the instructions for use!

The glass anti-roll plate can remain in place during disinfection.

- 1. Set the cryochamber temperature up to a value of -20 °C or above.
- 2. Remove the knife or blade from the knife/blade holder.
- 3. Remove all specimens, microscope slides and tools from the cryochamber.
- 4. Remove debris from the cryochamber.

Allow the cryochamber to reach the previously selected temperature.

Once the selected temperature is reached, either

5a. Spray the surfaces to be disinfected with a uniform coat of disinfectant (when using Leica Cryofect, a maximum of 10 sprays are allowed - see 2.6. "Cleaning and disinfection") - while doing so, the surfaces should be covered with a uniform coat of film - or

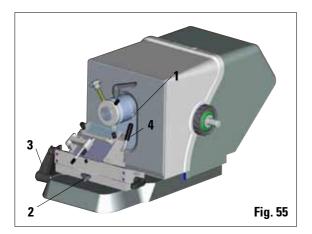
5b. soak a cloth with disinfectant and apply it on the contaminated surfaces.

- 6. In both cases, allow the disinfectant a reaction time of at least 15 minutes.
- 7. After the reaction time, wipe the disinfectant off using a tissue.
- 8. Dispose of the tissue according to applicable laboratory regulations.
- 9. Set the cryochamber temperature to the originally selected value.



If increased frost buildup occurs, start a manual defrost cycle.

9.3 Maintenance





9.3.1 General maintenance instructions

The microtome is virtually maintenance-free. To ensure a smooth operation of the instrument over several years we recommend the following:

- At least once a year, have the instrument inspected by a qualified service engineer authorized by Leica.
- Enter into a service contract at the end of the warranty period. For further information, please contact your local Leica service center.
- Clean the instrument every day.

Weekly:



Before the following parts can be oiled, all section waste and deposits must be carefully removed.

- Oil the plastic coupling (6, Detail 57a) with a drop of cryostat oil (Type 407);
- Lubricate the specimen cylinder (1):
 Push the appropriate coarse feed button to move the specimen cylinder out to the front stop position, apply a drop of cryostat oil and move the specimen cylinder back to the home position by pressing the appropriate coarse feed button.

Occasionally, or when required:

 Apply a drop of cryostat oil (Type 407) to the clamping piece (T-piece) (2, Fig. 55) on the microtome base plate and the clamping lever (3); Oiling the slot cover (4, Fig. 55).

To do so, first move the specimen head all the way upwards by turning the handwheel, apply a few drops of cryostat oil (Type 407) to the slot cover; then, move the specimen head all the way down and apply a few drops of cryostat oil.

Distribute the oil by turning the handwheel or using a clean tissue.

In case of visible pollution (such as dust), clean the air inlet opening (5, Fig. 56) of the condenser
on the bottom right-hand side of the instrument using a brush, broom or extraction cleaner in the
direction of the louvers.



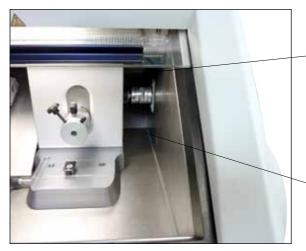
Do not carry out any repairs on your own as this will invalidate the warranty. Repairs may only be carried out by qualified service engineers authorized by Leica.



The germicidal UV lamp is a simple and safe means of minimizing pathogens, which considerably reduces the risk of infection.

Nevertheless, UV light disinfection cannot replace chemical disinfection of the cryochamber at regular intervals.

UV light disinfection is effective only on those surfaces that are directly illuminated.



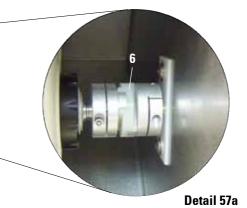


Fig. 57

9.3.2 Replacing the fuses



Turn the instrument off and disconnect the power plug before replacing the fuses! Use only the fuse types specified in Chapter "3. Technical Data.

Failure to comply with this can cause severe damage to the instrument and its surroundings and a risk of fatal injury!



Fig. 58

Protection	Туре
Display	T 0.25 A
Coarse feed	T 1.6 A
Processor board supply	T 1.0 A
Peltier element	T 6.25 A
Heaters	T 4 A
	Display Coarse feed Processor board supply Peltier element

On the back of the instrument there is a fuse box with 5 fuses:

- Unscrew the fuse cap with a screwdriver.
- Remove both fuse cap and fuse.
- Insert the defined fuse into the sleeve, then use the screwdriver to screw the sleeve back in as far as it will go.

9.3.3 Replacing the UVC lamp



Switch the instrument off and disconnect the power plug before replacing the fluorescent lamp! Wear suitable safety gloves and safety glasses.

If the lamp is broken, it must be replaced by the technical service, as the replacement involves a high risk of injury.

A UVC lamp has an estimated service life of approx. 9,000 hours.

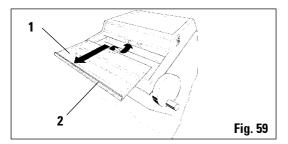
Each on/off switching cycle reduces the lamp life by approx. one hour plus burning time (30 minutes or 180 minutes respectively).



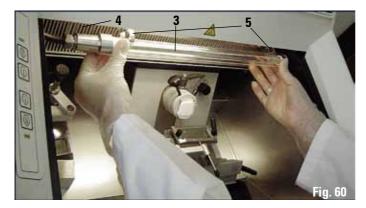


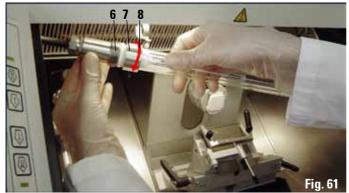


When both disinfection indicator lights (short and long-term disinfection) are flashing alternately in control panel field 1, the UVC lamp must be replaced.

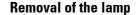


- Switch off the instrument at the circuit breaker.
- Disconnect the power plug.
- Slightly lift the sliding window (1) using the grip (2) and pull it out to the front.









The UVC lamp (3) is installed in front of the protection screen for chamber illumination (4).

- Hold the lamp with both hands and carefully pull it out of the clips (5) with a slight forward movement.
- Detach the metal ring (7) on the holder (6) in the direction of the arrow (8) and carefully pull the lamp out of the holder with your right hand (see Fig. 60 + 61).

Installation of the new lamp

- Carefully slide the metal ring (7) over the lamp from the left (see Fig. 62).
- Push the lamp into the holder on the left side until it engages.
- Screw the metal ring onto the holder, then hold the lamp with both hands and carefully push it into the clips (5).
- Replace the sliding window.
- Connect the instrument to the power supply again and switch it on.

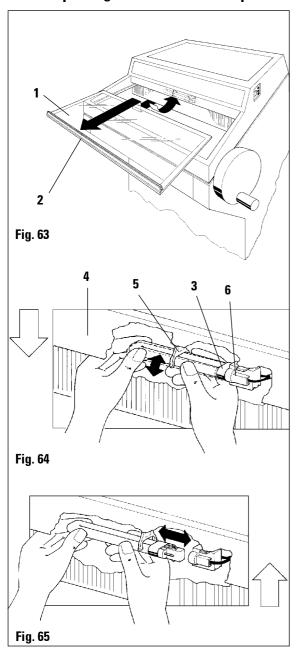


If any metallic mercury is released, handle it carefully and dispose of it properly!



Dispose of the UVC lamp separately!

9.3.4 Replacing the fluorescent lamp



- Switch off the instrument at the circuit breaker.
- Disconnect the power plug.
- Slightly lift the sliding window (1) holding it by the grip (2) and pull it out to the front.
- For lamp data, refer to Chapter 3, "Technical Data".



If the lamp breaks, it is mandatory to call Technical Service for replacement. RISK OF INJURY!

If any metallic mercury is released, handle it carefully and dispose of it properly!

Uninstalling the lamp (cryochamber illumination)

The fluorescent lamp (3) is installed hidden behind the protection screen (4).



Wear suitable safety gloves and safety glasses.

- Touch the lamp for better orientation.
- **Slightly** tilt the lamp down to the left and pull it out of the clip (5).
- Hold the lamp with both hands and pull it to the left out of the holder (6).

Installation of the new lamp

- Hold the lamp in the correct mounting position (Fig. 65) as shown and push it to the right until it engages in the holder.
- Push the lamp upwards slightly into the clip (5).
- Replace the sliding window.
- Connect the instrument to the power supply again and switch it on.

10.1 Ordering information

Blade/knife holder and blade/knife holder base

Diddo, Killio lioldol dila biddo, Killio lioldol base	
Blade/knife holder base for CN, CE	
Knife holder CN for standard microtome knife	
Knife holder CN-Z for TC microtome knife	
Blade holder CE, high-profile and low-profile	
Blade holder CE, without clearance angle adjustment	
Blade holder pressure plate, 22°, for microtome blades	
Blade holder CE-TC	14 0491 47874
Anti-static kit for blade holder CE, high-profile	14 0800 37740
Anti-static kit for blade holder CE, low-profile	14 0800 37739
Knife support for knife holder CN, for short knives	14 0419 19426
Knife support for knife holder CN, for long knives	14 0419 19427
Disposable blades	
Leica TC-65 disposable blades, 5 pcs	14 0216 26379
Leica high-profile disposable blades, Type 818, 1 pack of 50,	
Leica high-profile disposable blades, Type 818, 10 packs of 50,	
Leica low-profile disposable blades - Type 819, 1 pack of 50	
Leica low-profile disposable blades, Type 819, 10 packs of 50	
Reusable knives	
Knife, 16 cm length, profile c	14 0216 07100
Knife, 16 cm carbide metal blade, profile c	
Knife, 12 cm length, profile c	
Knife, 16 cm length, profile d	
Knife, 16 cm carbide metal blade, profile d	
Knife, 12 cm length, profile d	14 0216 07130
Variable knife case for 1 or 2 knives with length of 10 to 16 cm	14 0213 11140
Dr. Peters cryoembedding system -	
Intended use: enables accurate orientation of the tissue samples	
Case with cryoembedding system	14 0201 40670
Cryoembedding system	
Embedding well bar set with small recesses, 18 mm	
Embedding well bar set with medium-sized recesses, 24 mm	
Embedding well bar set with large recesses, 30 mm	
Freezing griddle/elevated freezing block set	
<u>.</u>	

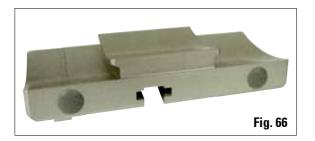
10. Ordering Information, Components and Consumables

Embedding well bar, 4 x 18 mm	
Embedding well bar, 4 x 24 mm	
Embedding well bar, 3 x 30 mm	
Embedding well bar, 2 x 20 x 6 mm, 2 x 20 x 9 mm	
Embedding well bar, 2 x 24 x 6 mm, 2 x 24 x 9 mm	
Specimen stage, rectangular, 28 mm	14 0201 39123
Specimen stage, rectangular, 36 mm	14 0201 39124
Heat extractor	14 0201 39125
Container for specimen stages	
Dispensing slides for Dr. Peters' cryoembedding system, pack of 8	14 0201 39127
Components and consumables	
Intended use: Enables manual staining of cryosections	
Easy Dip staining container, white	14 0712 40150
Easy Dip staining container, pink	
Easy Dip staining container green	
Easy Dip staining container, yellow	
Easy Dip staining container, blue	
Easy Dip staining rack, gray	
Blue O-ring, for color marking, 20 mm and 30 mm, 10 pcs.	
Red O-ring, for color marking, 20 mm and 30 mm, 10 pcs.	
Blue O-ring, for color marking, 40 mm, 10 pcs	
Red O-ring, for color marking, 40 mm, 10 pcs.	
Blue O-ring, for color marking, 55 mm, 10 pcs.	
Red O-ring, for color marking, 55 mm, 10 pcs.	
Toolset for cryostats	
Specimen disc, 20 mm	
Specimen disc, 25 mm	
Specimen disc, 30 mm	
Specimen disc, 40 mm	14 0370 08637
Specimen disc, 55 mm	
Specimen disc, 50 x 80 mm	
Intended use: speeds up the process of freezing tissue samples	
Heat extractor – stationary, assembly	14 0471 30792
Parking station	
Heat extractor - mobile	
Transfer block for specimen discs, small (holder for specimen discs)	
Transfer block for opcomion discs, small (notice for specimen discs)	17 0701 7/701

10. Ordering Information, Components and Consumables

Transfer block for specimen discs, large (holder for specimen discs) Thermal block Intended use: helps to warm up the specimen if too cold or	14 0491 47786
pick up specimen from specimen disc	1/ 0202 125/2
Miles adapter, for TissueTek specimen discs	
Plug adapter set EU-UK	
Shelf, movable, assembly	
Sileil, illuvable, assembly	14 0431 40730
Consumables	
Add-on kit: Swing arm with anti-roll guide system	
Anti-roll guide system CE, glass - 70 mm, 100 μm spacer for 5 - 50 μm	
Anti-roll guide system CE, glass - 70 mm, 50 μm spacer for up to 4 μm	
Anti-roll guide system CE, glass - 70 mm, 150 μm spacer for over 50 μm	
Anti-roll guide system for knife holder CN or CN-Z, for 5 µm - 50 µm	14 0419 33981
Glass insert, glare-reducing, 70 mm wide	14 0477 42497
Glass insert - 50 mm	
Embedding medium for cryosectioning "Jung tissue freezing medium", 125 ml	14 0201 08926
Cryostat oil - Type 407, 250 ml	
Leica Cryofect - low-temperature disinfectant, 4 x 350 ml	
Safety gloves, cut-resistant, size S	
Safety gloves, cut-resistant, size M	14 0340 29011
UV fluorescent tube	14 04714 0422
Components and consumables for manual knife grinding	
Lapping device for knife, profile c, length 16 cm	14 0337 08687
Lapping device for knife, profile c, length 12 cm	
Lapping device for knife, profile c, length 10 cm	14 0337 08689
Lapping device for knife, profile d, length 16 cm	14 0337 08693
Handle for knives from 10 cm to 30 cm in length	14 0283 08504
Lapping stone (fine), yellow with dark back, approx. 25 x 5.5 cm	14 0210 04070
Lapping stone (coarse), blue-green, with scraper, approx. 25 x 5.5 cm	14 0210 04071
Metal case for storing lapping stones	
Strop, Type "Heidelberg"	
Screw clamp for fastening strop, Type "Heidelberg"	14 0285 07057
Strop paste, red	14 0209 04064

10. Ordering Information, Components and Consumables



Knife/blade holder base.

for knife holder CN, blade holder CE

Order No.14 0491 47875



Knife holder CN,

for standard microtome knives or magnetic blade rails.

Clearance angle adjustment and knife height adjustment.

Support rails for long and short knives.

Anti-roll guide and adjustable knife guard.

Order No.14 0477 42358

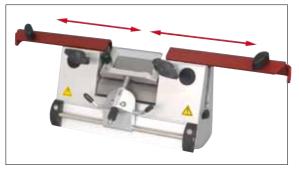


Fig. 68

Knife holder CN-Z,

For conventional and tungsten carbide knives up to 16 cm in length. Knife pressure plate for extreme stability and full utilization of the knife blade.

Clearance angle adjustment and knife height adjustment, anti-roll guide and adjustable knife guard.

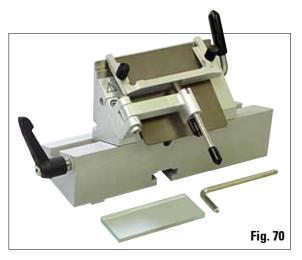
Order No.14 0477 42363



Blade holder CE, universal (high-profile and low-profile disposable blades)

with lateral movement and glass anti-roll guide. Adjustable clearance angle.

Order No.14 0491 47873



Blade holder CE.

for low-profile disposable blades, **without** clearance angle adjustment,

including blade/knife holder base and pressure plates

Order No.14 0419 33992



Fig. 71

Blade holder CE-TC,

for carbide metal disposable blades. Particularly suitable for sectioning harder materials, such as tissue, bone or cartilage.

Order No.14 0491 47874



Blade holder pressure plate, 22°, for high-profile microtome blades Order No.14 0491 48004

Fig. 72



Anti-static kit, blade holder CE
Order No.14 0800 37740

Fig. 73

Fig. 74



Order No.14 0419 19426

Knife support for knife holder CN, for long knives

Order No.14 0419 19427

Knife support for knife holder CN,

for short knives



Disposable blades Leica TC-65,

Leica TC-65 microtome, disposable blade system for sectioning hard specimen materials.

The Leica TC-65 carbide metal disposable blades were specially developed for the requirements in labs where hard, blunt materials are routinely sectioned.

The one-of-a-kind fine-grain carbide metal guarantees sections to approx. 1 μm . The blades are fully recyclable.

Length: 65 mm, thickness: 1 mm, height: 11 mm Leica TC-65 carbide metal disposable blades. Pack of 5.

Order No.14 0216 26379



Leica high-profile disposable blades, Type 818 $80 \times 14 \times 0.317 \ mm$





Fig. 77

Leica low-profile disposable blades, Type 819 $80 \times 8 \times 0.25$ mm

1 pack of 50	. 14	0358	38925
10 packs of 50	. 14	0358	38382



Knife, 16 cm length, profile c

(flat on both sides, for wax and frozen sections) Note: including knife case 14 0213 11140

Order No.14 0216 07100



Knife, 16 cm carbide metal blade, profile c

Note: including knife case 14 0213 11140

Order No.14 0216 04206

Knife, 12 cm length, profile c

Note: including knife case, 14 0213 11140

Order No.14 0216 07092





Knife, 16 cm length, profile d,

Note: including knife case 14 0213 11140

Order No.14 0216 07132



Knife, 16 cm carbide metal blade, profile d,

Note: including knife case

14 0213 11140

Order No. 14 0216 04813



Knife, 12 cm length, profile d,

for Leica CM1500 / CM1800

Note: including knife case 14 0213 11140

Order No.14 0216 07130

Fig. 81



Variable knife case,

for 1 to 2 knives from 10 to 16 cm in length

Order No.14 0213 11140

Variable knife case,

for 1 or 2 knives with length of 18.5, 22 or 30 cm (for carbide metal or SM2500 knives: only for 1 knife!) only for 1 knife!)

Order No.14 0213 11141



Case with cryoembedding system includes:

- 3 embedding well bars with recesses, in three sizes: 18 mm, 24 mm, 30 mm
- 6 specimen stages, small
- 4 specimen stages, large
- 4 heat extractors
- 1 container for specimen stages
- 16 dispensing slides
- 1 sectioning board/freezing griddle
- 1 heat extractor, elevated
- 1 embedding forceps, angled

Order No.14 0201 40670



Fig. 84



Fig. 85

Cryoembedding system, complete

consisting of:

- 3 embedding well bars with recesses, in three sizes: 18 mm, 24 mm, 30 mm
- 6 specimen stages, small
- 4 specimen stages, large
- 4 heat extractors
- 1 container for specimen stages
- 16 dispensing slides
- 1 sectioning board/freezing griddle
- 1 heat extractor, elevated
- 1 embedding forceps, angled

Order No.14 0201 39115

Embedding well bar set with small recesses consisting of:

- 1 embedding well bar, recesses 18 mm
- 4 specimen stages, small
- 2 heat extractors
- 8 dispensing slides

Order No.14 0201 39116

Embedding well bar set with medium-sized recesses consisting of:

- 1 embedding well bar, recesses 24 mm
- 4 specimen stages, small
- 2 heat extractors
- 8 dispensing slides

Order No.14 0201 39117

Embedding well bar set with large recesses consisting of:

- 1 embedding well bar, recesses 30 mm
- 4 specimen stages, large
- 2 heat extractors
- 8 dispensing slides

Order No.14 0201 39118



Fig. 86

Set of freezing griddle/heat extractor,

elevated consisting of:

- 1 forceps, epoxy-coated
- 1 putty knife
- Extra elevating bars

Order No.14 0201 39119

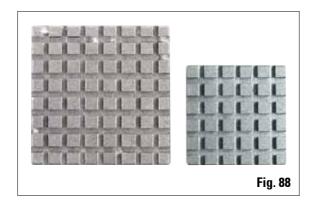


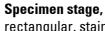
Fig. 87

Embedding well bar

Hoight: 25 1 mm

Height: 25.4 mm
with 4 recesses, size 18 mm, stainless steel
Order No14 0201 39120
Height: 25.4 mm
with 4 recesses, size 24 mm, stainless steel
Order No14 0201 39121
Height: 25.4 mm
with 4 recesses, size 30 mm, stainless steel
Order No14 0201 39122
Height: 25.4 mm
with 2 recesses, size 20 x 6 mm, stainless steel
with 2 recesses, size 20 x 9 mm, stainless steel
Order No14 0201 46662
Height: 25.4 mm
with 2 recesses, size 24 x 6 mm, stainless steel
with 2 recesses, size 24 x 9 mm, stainless steel
Order No14 0201 46663





rectangular, stainless steel, small - 28 mm Pack of 1.

Order No.14 0201 39123

Specimen stage,

rectangular, stainless steel, large - 36 mm Pack of 1.

Order No.14 0201 39124



Order No.14 0201 39125



Container for specimen stages

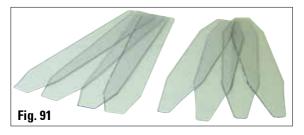
Order No.14 0201 39126



Dispensing slide for Dr. Peter's cryoembedding system

pack of 8.

Order No.14 0201 39127





Blue O-ring,

for color marking of the specimen disc 20 and 30 mm (14 0477 40044) pack of 10.

Order No.14 0477 43247

Red O-ring,

for color-marking the 20 mm and 30 mm specimen disc (14 0477 40044)

pack of 10.

Order No.14 0477 43248

Fig. 92 Blue O-ring,

for color-marking the 40 mm specimen disc (14 0477 40045)

pack of 10.

Red O-ring,

for color-marking the 40 mm specimen disc (14 0477 40045)

pack of 10.

Order No.14 0477 43250

Blue O-ring,

for color-marking the 55 mm specimen disc (14 0477 40046)

pack of 10.

Order No.14 0477 43251

Red O-ring,

for color-marking the 55 mm specimen disc (14 0477 40046)

pack of 10.

Order No.14 0477 43252



Fig. 93

Toolset for cryostats,

consisting of:

00	moloting on	
-	1 brush, narrow	14 0183 28642
-	1 Leica brush with magnet	14 0183 40426
-	1 key with handle, No. 5	14 0194 04760
-	1 Allen key, No. 2.5	14 0222 04137
-	1 Allen key, No. 3.0	14 0222 04138
-	1 Allen key, No. 4.0	14 0222 04139
-	1 Allen key, No. 5.0	14 0222 04140
-	1 Allen key, No. 6.0	14 0222 04141
-	1 Allen key, No. 1.5	14 0222 10050
-	1 Allen key, ball-cage, No. 4	14 0222 32131
-	1 single-head wrench, No. 16	14 0330 18595
-	1 single-head wrench, No. 13	14 0330 33149
	,	

Order No.14 0436 43463



Easy Dip, specimen slide holder,

for 12 specimen slides, gray pack of 6

Order No.14 0712 40161



Fig. 95

Easy Dip, staining container Color: white
pack of 6
Order No14 0712 40150
Color: pink
pack of 6
Order No14 0712 40151
Color: green
pack of 6
Order No14 0712 40152
Color: Yellow
pack of 6
Order No14 0712 40153
Color: Blue
pack of 6
Order No14 0712 40154



Specimen disc,

20 mm	Order No. 14 0370 08636
25 mm	14 0416 19275
30 mm	14 0370 08587
40 mm	14 0370 08637
55 mm	14 0419 26491
50 v 80 mm	1/1 0/19 26750

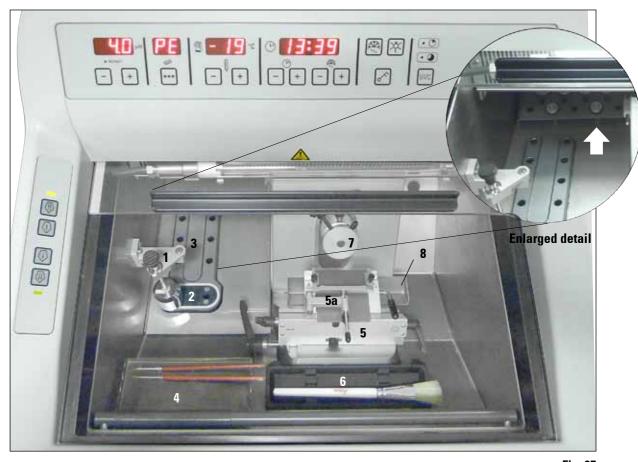
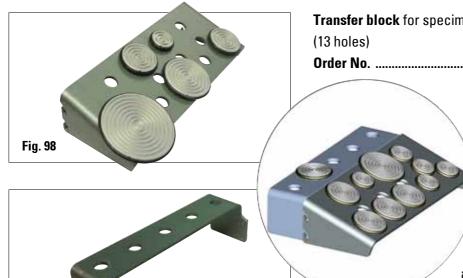


Fig. 97

- 1 Heat extractor, stationary (optional)
- 2 Peltier element (with 2 stations)
- 3 Freeze shelf, 8 positions
- 4 Shelf, movable (optional)
- 5 Blade holder CE with anti-roll guide (a) antiroll guide also acts as knife guard (optional)
- 6 Brush shelf

- 7 Object head, directional
- 8 Waste tray

Enlarged detail - Storage space for both transfer blocks



Transfer block for specimen discs, large

Order No.14 049147786

Transfer block for specimen discs, small

(5 holes) - combinable with transfer block for specimen discs, large

Order No.14 0491 47787



Heat extractor, mobile (see also page 87)



Thermal block,

for easy removal of frozen blocks from the specimen disc (see also page 91)

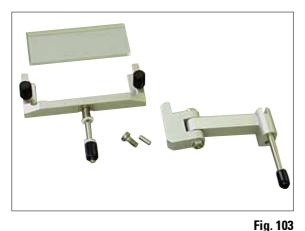
Order No.14 0398 18542

Fig. 99



Adapter,

for Miles Tissue Tek specimen discs Order No.14 0436 26747



Add-on kit:

Swing arm with anti-roll guide system

consisting of:

Swing arm

Anti-roll guide system:

- Glass insert 70 mm
- Metal frame for exchangeable glass inserts CE
- 100 µm spacer
- Recommended section thickness 5 μ m-50 μ m

For blade holder CE:

(14 0491 47873, 14 0419 33992)

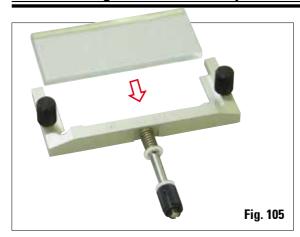
Fig. 104

Anti-roll guide system CE, glass - 70 mm consisting of:

- Glass insert 70 mm
- Metal frame for exchangeable glass inserts CE
- 100 µm spacer
- Recommended section thickness 5 µm-50 µm

For blade holder CE: (14 0491 47873, 14 0419 33992)

Order No.14 0419 33980



Anti-roll guide system CE, glass - 70 mm

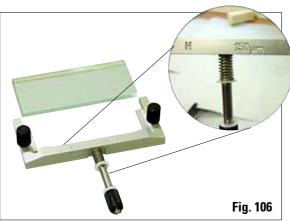
For specific application

consisting of:

- Glass insert 70 mm
- Metal frame for exchangeable glass inserts CE
- 50 µm spacer
- Recommended section thickness up to 4 µm

For blade holder CE: (14 0491 47873, 14 0419 33992)

Order No.14 0419 37258



Anti-roll guide system CE, glass - 70 mm

For specific application consisting of:

- Glass insert 70 mm
- Metal frame for exchangeable glass inserts CE
- 150 µm spacer
- Recommended section thickness over 50 µm

For blade holder CE:

(14 0491 47873, 14 0419 33992)

Order No.14 0419 37260



Anti-roll guide system,

for knife holder CN or CNZ glass - 50 mm consisting of:

- Glass insert 50 mm
- Metal frame for exchangeable glass inserts CN, CNZ
- Recommended section thickness 5 μm 50 μm

Order No.14 0419 33981



Glass plate,

glare-reducing, 70 mm wide Spare glass for anti-roll guide systems CE-BB: (14 0477 42491, 14 0477 42492, 14 0477 42493) for blade holder CE-BB (14 0477 43005)

Order No.14 0477 42497



Glass insert - 50 mm

For knife holder CN (14 0477 42358)

Order No.14 0419 33816



Embedding medium,

for cryosectioning

"Jung tissue freezing medium," 125 ml

Order No.14 0201 08926



Cryostat oil,

- Type 407, 250 ml bottle

Order No.14 0336 06100



Caution: Dangerous goods!

Leica Cryofect,

Low-temperature disinfectant, 4 350 ml bottles.

For temperatures down to -20 °C.

Order No.14 0419 33816



Cut-resistant safety gloves,

1 pair, size S

Order No.14 0340 40859

Cut-resistant safety gloves,

1 pair, size M

Order No.14 0340 29011



.

Heat extractor, stationary, assembly Order No.14 0471 30792

Low-temperature buffer for heat extractor ("parking station") 14 0471 30793

Fig. 114

Fig. 113

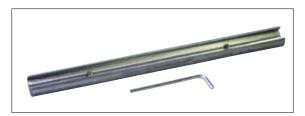


Fig. 115

Lapping device for knife,

Profile c, 16 cm length

Order No.14 0337 08687

Lapping device for knife,

Profile c, length: 12 cm

Order No.14 0337 08688

Lapping device for knife,

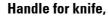
Profile c, 10 cm length

Order No.14 0337 08689

Lapping device for knife,

Profile d, length: 16 cm

Order No.14 0337 08693



length from 10 cm to 30 cm

Order No.14 0283 08504



Fig. 116



Lapping stone (fine),

Yellow with dark back, approx. 25 x 5.5 cm **Order No.14 0210 04070**

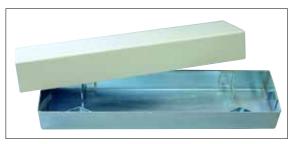
Fig. 117



Lapping stone (coarse),

blue-green, with scraper, approx. 25 x $5.5\,\mathrm{cm}$

Order No.14 0210 04071



Metal case,

for storing lapping stones Order No. 14 0337 04070 or 14 0337 04071

Order No.14 0210 04073

Fig. 119



Fig. 120

Strop, Type "Heidelberg"

Order No.14 0209 04066



Screw clamp,

for fastening strop of type "Heidelberg"

Order No.14 0285 07057

Fig. 121



Strop paste, red

Order No.14 0209 04064

Fig. 122



UV fluorescent tube

Order No.14 047140422



Storage system, movable

for installation in the front part of the cryostat for cooled storage of preparation aids

Order No.14 0491 46750

10.2 Mobile heat extractor – application



Fig. 125

Specimen freezing with the freezing shelf can be accelerated by the additional use of a heat extractor.

- Store the heat extractor in the cryochamber.
- Place it on the specimen surface to accelerate freezing.
- Remove it once the specimen is entirely frozen.

Recommendation:



It is recommended to precool the heat extractor in liquid nitrogen or other refrigerant.

10.3 Cold extractor – application

The thermal block (8) facilitates the removal of the frozen specimen from the specimen disc.



Keep the thermal block outside the cryochamber at room temperature. Place it in the cryochamber only for specimen removal.



- Place the cap (9) on the required side, so that the appropriate location hole for the specimen disc is visible.
- Fit the pin (6) of the specimen disc (5) into the corresponding bore (7) on the cold extractor.
- After about 20 seconds, the frozen specimen can be removed from the specimen disc with forceps.
- If the cap is too loose, readjust it with the small screw (10).
 - Do not overtighten the screw.
- After removing the specimen, remove the cold extractor from the cold cryochamber once again.

Warranty

Leica Biosystems Nussloch GmbH guarantees that the contractual product delivered has been subjected to a comprehensive quality control procedure based on the Leica in-house testing standards, and that the product is faultless and complies with all technical specifications and/or agreed characteristics warranted.

The scope of the warranty is based on the content of the concluded agreement. The warranty terms of your Leica sales organization or the organization from which you have purchased the contractual product shall apply exclusively.

Service information

If you are in need of technical customer support or spare parts, please contact your Leica representative or the Leica dealer where you purchased the instrument.

Please provide the following information:

- Model name and serial number of the instrument.
- Location of the instrument and name of a contact person.
- Reason for the service call.
- Delivery date

Decommissioning and disposal

The instrument or parts of the instrument must be disposed of according to existing applicable, local regulations.

Dispose of UV tube separately.

Dear Customer,

Any product that is to be returned to Leica Biosystems or serviced on site must be cleaned and decontaminated in the appropriate manner. Since it is not possible to decontaminate for prion diseases, such as CJD, BSE, CWD etc., equipment exposed to specimens containing prion diseases cannot be returned to Leica Biosystems for repair. Prion-contaminated instruments will only be repaired after the service technician has been made aware of the potential risks, informed about the applicable directives and procedures for the respective institution, and provided with personal protective equipment. Please complete this certificate carefully and place a copy with the instrument, attach it to the outside of the transport case or hand it directly to the service engineer. If packages are returned, they will not be opened or have maintenance measures begun until the Company or service technician has received confirmation of decontamination. Should returned goods be considered a hazard by the Company, they will be returned immediately to the customer at his/her expense. Note: Microtome knives must be packed in the corresponding box.

Des	cription				
		Name/Model		Fabr. No.	
		KAT No.		Quantity	
		Check box A if applicable. (se please complete all parts of B, provid- or appropriate.	
Α	Yes	This equipment has not been in contact with unfixed biological samples.			
B 1 Yes No		Internal or external areas of the instrument have been exposed to the following hazard- ous substances: Blood, body fluids, pathological samples Other biohazards Chemicals/substances hazardous to health Substances Radioactivity Other hazards This equipment has been cleaned and decontaminated:			
	Yes No	If yes, give details of the method:		Additional information	
		If no*, please indicate why not:			
		* Such equipment must not be returned without	out the wr	ritten agreement of Leica Biosystems.	

3				
Yes No The equipment has been prepared to ensure safe handling/transportation. Whenever possible, please use the original packaging.				
Important - to avoid refusal of shipment: Place one copy in the unit prior to packaging sumes all responsibility for the immediate redecontamination documentation.				
If you have any further questions, please call	your local Leica branch	n office.		
Leica Internal Use: If applicable, note corresp	onding Job and RAN/R	GA numbers:		
Job Sheet No.:				
SU Return Goods Authorization:				
BU R eturn A uthorization N umber:	<u>-</u>			
Signature/Date	Institute			
	Department			
Name				
	Address			
Position				
E-mail	Phone	Facsimile		



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